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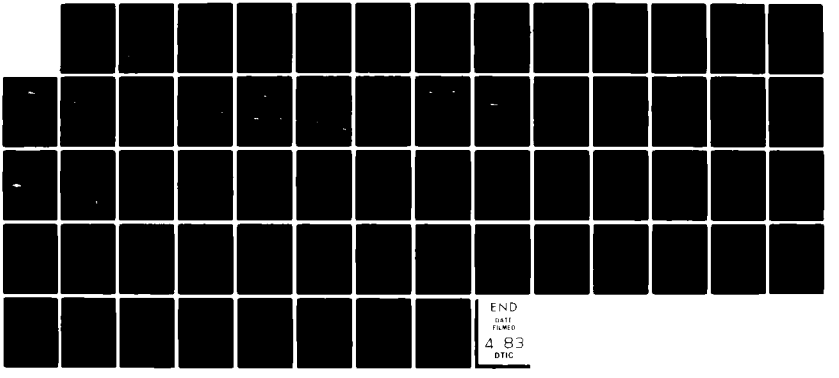

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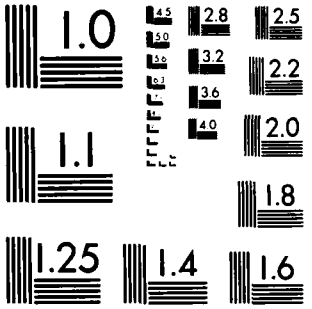

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NATIONAL BUREAU OF STANDARDS-1963-A

STUDIES OF PHLEBOTOMINE SAND FLIES

ANNUAL REPORT

By

D. G. YOUNG

31 August 1980

Supported by

U. S. ARMY MEDICAL RESEARCH & DEVELOPMENT COMMAND
Fort Detrick, Frederick, Maryland 21701

CONTRACT DADA-17-72-C-2139

DEPARTMENT OF ENTOMOLOGY & NEMATOLOGY
UNIVERSITY OF FLORIDA
Gainesville, Florida 32611

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SECURITY CLASSIFICATION OF THIS PAGE (When Data Entered)

REPORT DOCUMENTATION PAGE	READ INSTRUCTIONS BEFORE COMPLETING FORM
1. REPORT NUMBER / 2. GOVT ACCESSION NO. AD-A126733	3. RECIPIENT'S CATALOG NUMBER
4. TITLE (and Subtitle) STUDIES OF PHLEBOTOMINE SAND FLIES	5. TYPE OF REPORT & PERIOD COVERED Annual 31 August 1979-31 August 1980
	6. PERFORMING ORG. REPORT NUMBER
7. AUTHOR(s) D.G. Young	8. CONTRACT OR GRANT NUMBER(s) DADA 17-72-C-2139
9. PERFORMING ORGANIZATION NAME AND ADDRESS University of Florida Gainesville, Florida 32611	10. PROGRAM ELEMENT, PROJECT, TASK AREA & WORK UNIT NUMBERS 62759A 3A762759A831.00.031
11. CONTROLLING OFFICE NAME AND ADDRESS U.S. Army Medical Research and Development Command Fort Detrick, Frederick, MD 21701	12. REPORT DATE 31 August 1980
	13. NUMBER OF PAGES ~~56~~ 62
14. MONITORING AGENCY NAME & ADDRESS(if different from Controlling Office)	15. SECURITY CLASS. (of this report) unclassified
	15a. DECLASSIFICATION/DOWNGRADING SCHEDULE

16. DISTRIBUTION STATEMENT (of this Report)

Approved for public release; distribution unlimited.

17. DISTRIBUTION STATEMENT (of the abstract entered in Block 20, if different from Report)

18. SUPPLEMENTARY NOTES

19. KEY WORDS (Continue on reverse side if necessary and identify by block number)

Lutzomyia
Phlebotominae
Phlebotomus
Leishmaniasis
Sand fly

20. ABSTRACT (Continue on reverse side if necessary and identify by block number)

A preliminary key to the phlebotomine sand flies of Kenya was prepared as an aid to identification. The need for fresh material became readily apparent. Further progress was made on illustrating important features of New World species, necessary for inclusion in a forthcoming handbook of the American sand flies. New geographic records and specimens for the reference collection were given to the Principal Investigator by colleagues working in Guatemala, Mexico and Brazil. Several descriptive papers were submitted for

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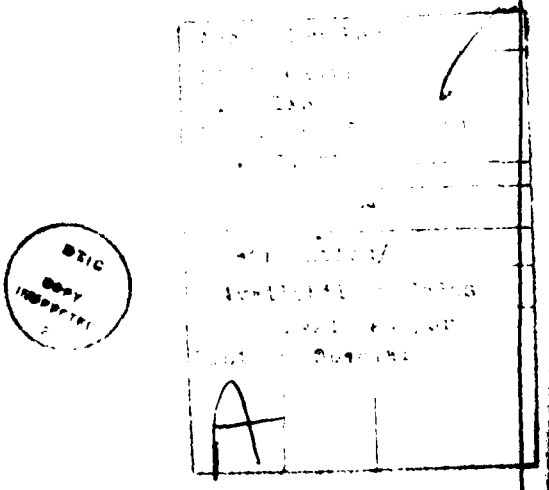

ABSTRACT

1. Preparing Institution: University of Florida
2. Title of Report: Studies of phlebotomine sand flies
3. Principal Investigator: David G. Young, Ph.D.
4. No. of Pages and Date: 56; 31 Aug. 1980
5. Contract No. DADA 17-72-C-2139
6. Supported by: Dept. of the Army, Washington, D.C. 20314

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7. Key Words: Sand fly

Lutzomyia

Phlebotominae

Phlebotomus

Leishmaniasis

Note: Copies of this report are filed with the Defense Document Center, Cameron Station, Alexandria, Virginia 33214, and may be obtained by qualified investigators working under government contract.

CONTENTS

PROGRESS REPORT

DADA 17-72-C-2139

Introduction

The importance of phlebotomine sand flies as vectors of leishmaniasis and arboviruses in many areas of the world is considerable. The W.H.O. scientific working group on leishmaniasis (1977 report) stated that ". . . taxonomic studies of these vectors are of fundamental importance due to the difficulty in identifying them." Recommendations of this group included continued maintenance of phlebotomine reference collections and support for research programs dealing with systematics of these insects. The group stressed the "need for new practical field keys" in some areas.

Objectives

The objectives under this contract are similar to those outlined by the W.H.O. working group. In addition to studying the American species (the projected date of completion of a handbook on these is June, 1982), we began to study the phlebotomines of Africa, beginning with Kenya, and the Middle East. Objectives include:

1. Preparing keys, illustrations, and other aids to identification both by geographic areas and by taxonomic groups.
2. Arriving at a more satisfactory classification of the subfamily phlebotominae.
3. Building a reference collection on a worldwide basis.
4. Maintaining one or more species in laboratory colonies to provide

immature stages and adults necessary for the evaluation of non-morphological taxonomic techniques.

Results

Slide mounted specimens of Kenyan phlebotomines (27 species) in our collection enabled the Principal Investigator and Capt. Raymond Beach to compile a preliminary identification key to the species known to occur there (Appendix I). The specimens on hand were poorly mounted over 20 years ago in temporary medium; therefore, it was not possible to critically evaluate all of the taxonomic characters believed to be important. A forthcoming trip to that country during June-July, 1981, working in cooperation with investigators at the International Centre of Insect Physiology and Ecology, Nairobi, as an invited scientist and WRAIR should be fruitful in terms of obtaining adequate samples of various species. One of the unsolved taxonomic problems of direct importance to the epidemiology of visceral leishmaniasis in Kenya is the apparent impossibility of separating the 3 *Synphlebotomus* spp. females by conventional means. One of these, *Phlebotomus martini* is the suspected vector but *P. celiae* may also be involved.

Work continued on the handbook of the New World species. The emphasis is being placed on identification; thus, many illustrations will be included--a time consuming but necessary aspect of the project. Most of the time spent on this handbook during the past year involved making drawings.

Colleagues sent specimens to the PI from Mexico, Guatemala, Brazil and the U.S.A. for determination. About 1000 flies were slide mounted and identified; those from Mexico and Guatemala, both little-collected

areas, were especially interesting. Dr. Charles Porter, CDC medical entomologist in Guatemala, collected 9 species previously unknown in the Republic. These are *Brumptomyia hamata, B. galindoi, Lutzomyia odax, L. ovallesi, L. carpenteri, L. shannoni, L. texana, L. trinidadensis* and *L. panamensis*. The latter species is an incriminated vector of cutaneous leishmaniasis in Panama.

Mexican material, collected at Miacatlán (Morelos) by Dr. Marco Camino and sent to the PI, is valuable because this locality is close to the type locality of 3 poorly known taxa--*L. durani, L. hardisoni* and *L. dodgei*. These species and at least 7 others, including the common *L. longipalpis*, were added to our reference collection. Information on these species will be included in the handbook.

Descriptions of two Amazonian species were sent to press (Appendices II and III). One of these forms, *L. olmeca nociva*--a man biter, belongs in the *flaviscutellata* complex which includes all the known vectors of *Leishmania mexicana* from Mexico to Brazil. The review of the *Lutzomyia davisi* complex should be published by April, 1981. A paper on a new group, the *Microps* group with 5 South American Species, was nearly completed.

Lutzomyia shannoni, a man-biting species in parts of its widespread geographic range (U.S.A.-Argentina), was collected for the first time in South Carolina and at Ft. Bragg, North Carolina, by LTC Ron Intermill in 1980. The identifications were made by the PI and CPT Peter Perkins, Ph.D. graduate student working with the PI [the fact that *shannoni* occurs at Ft. Bragg where *Leishmania*-infected soldiers apparently live and train indicates that transmission from vector to man or other mammal could occur]. The recent identification of *Leishmania infantum*, an Old World visceral

Leishmania, in Oklahoma in dogs (LTC Larry Hendricks, personal comm.), indicates that the successful establishment of an imported infection in the U.S.A. is possible. *L. shannoni* and two other U.S. species were successfully reared and colonized in our laboratory, mainly by graduate students working with the PI. Better methods of rearing resulted in a scientific note which was submitted to the Journal of Medical Entomology in 1980 (Appendix IV). The importance of rearing these insects in relation to taxonomy is clearly evident when morphologically inseparable, sibling species are considered. These methods for rearing immatures and handling adults of the *Synphlebotomus* complex in Kenya will be tried. If successful, then males and females in this complex can be properly associated, and identified specimens can be studied by various techniques, including cuticular hydrocarbon analysis.

Specimens of *L. anthophora* were sent to the Department of Arboviral Entomology, Ft. Detrick, for subcolonization and subsequent vector competence studies. The importance of maintaining phlebotomine colonies, especially vectors, is considerable--not only for taxonomic studies but also for experimental work not covered under this contract.

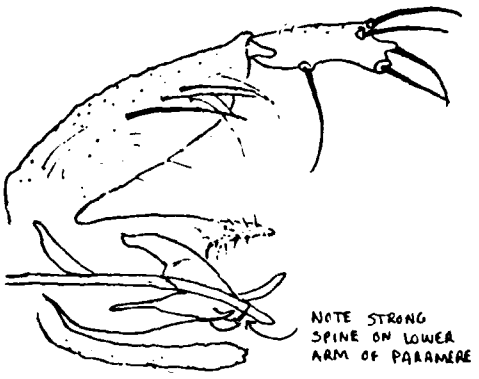
Appendix I

PRELIMINARY KEY TO THE PHLEBOTOMINAE OF KENYA

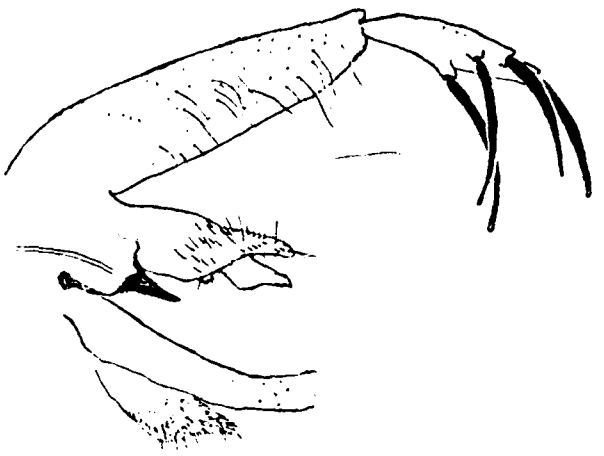
Males

1. Style with 5 major spines 2

   Style with 4 major spines 10



P. heischi



P. rodhaini

2. Paramere bilobed, i.e. forked. Pleural setae present..... S. heischi*

   Paramere simple. Pleural setae absent. (Genus Phlebotomus, in part). . 3

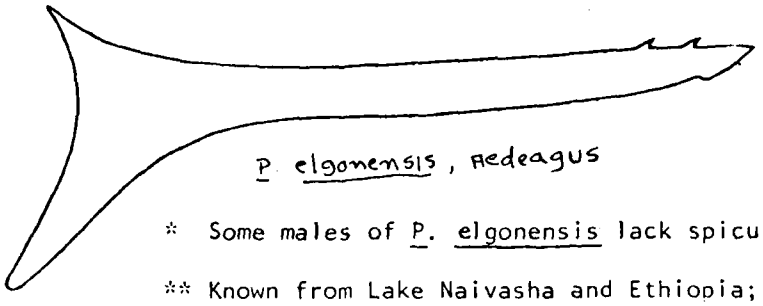
3. Genital filaments 3x to 11x length of pump. Persistent hairs inserted directly on coxite, i.e. no arm or tubercle. (Subgenus Larroussius) 4

   Genital filaments shorter than 3x length of pump. Coxite tuft of persistent hairs inserted on long arm. (Subgenus Synphlebotomus) . . . 8

* SEE COUPLET 6 OF FEMALE KEY FOR FIGURE OF PLEURAL SETAE.

4. Aedeagus with 1 or more subterminal spicules* 5

   Aedeagus without such spicules. 6



P. elgonensis, aedeagus

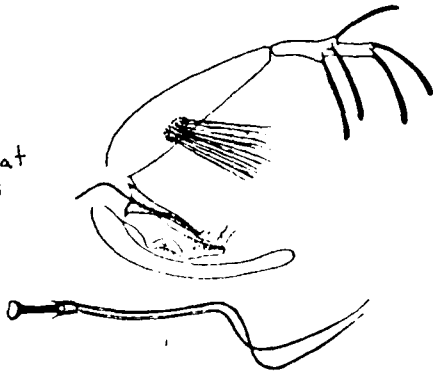
P. aculeatus

* Some males of P. elgonensis lack spicules.

** Known from Lake Naivasha and Ethiopia; elgonensis known only from caves on western slope of Mt. Elgon (2100 M to 2500 M).

5. 2/III-VII, 1/VIII-XV. Wing length 2.9-3.0 . . . . . P. aculeatus**

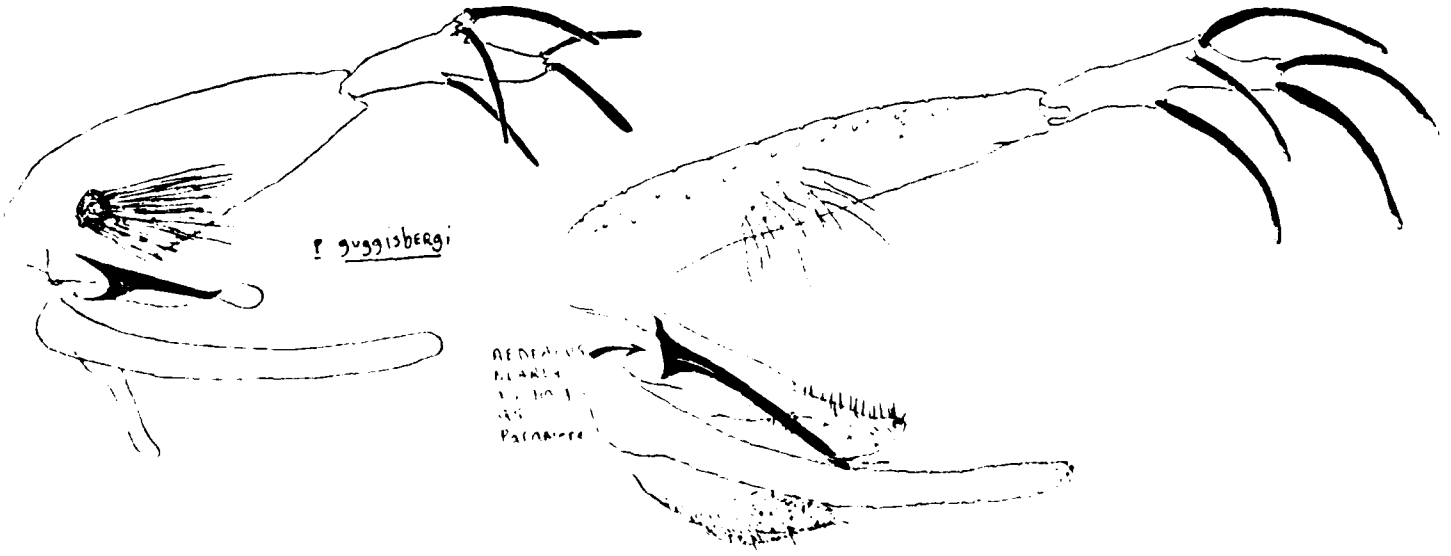
   2/III-XV. Wing length 3.2 - 3.44 . . . . . P. elgonensis



GENITALIA OF aculeatus; that OF elgonensis LOOKS VERY MUCH LIKE IT.

6. Coxite setae inserted on raspberry-like patch . . . . . P. guggisbergi

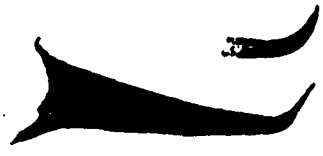
   Coxite setae more scattered, not inserted on a slightly raised patch. . 7



P. guggisbergi

7. Aedeagus nearly as long as parameres, its tip straight or nearly so. . . . . P. langeroni orientalis*

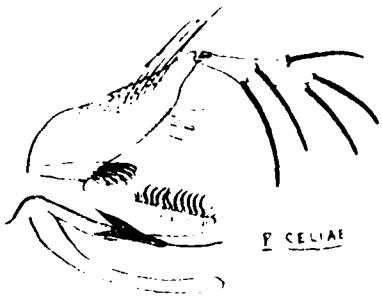
Aedeagus shorter than parameres, its tip turned upwards . . . P. pedifer**



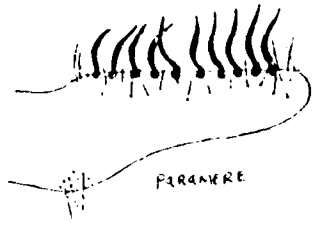
AEDEAGUS OF P. PEDIFER - UPTURNED AT TIP

8. Coxite setae of 7 or so spatulate hairs and some thin ones. Parameres with dorsal sickle-shaped setae. . . . . P. celiae

Coxite and paramere setae simple 9



P CELIAE



P celiae

PARAMERE

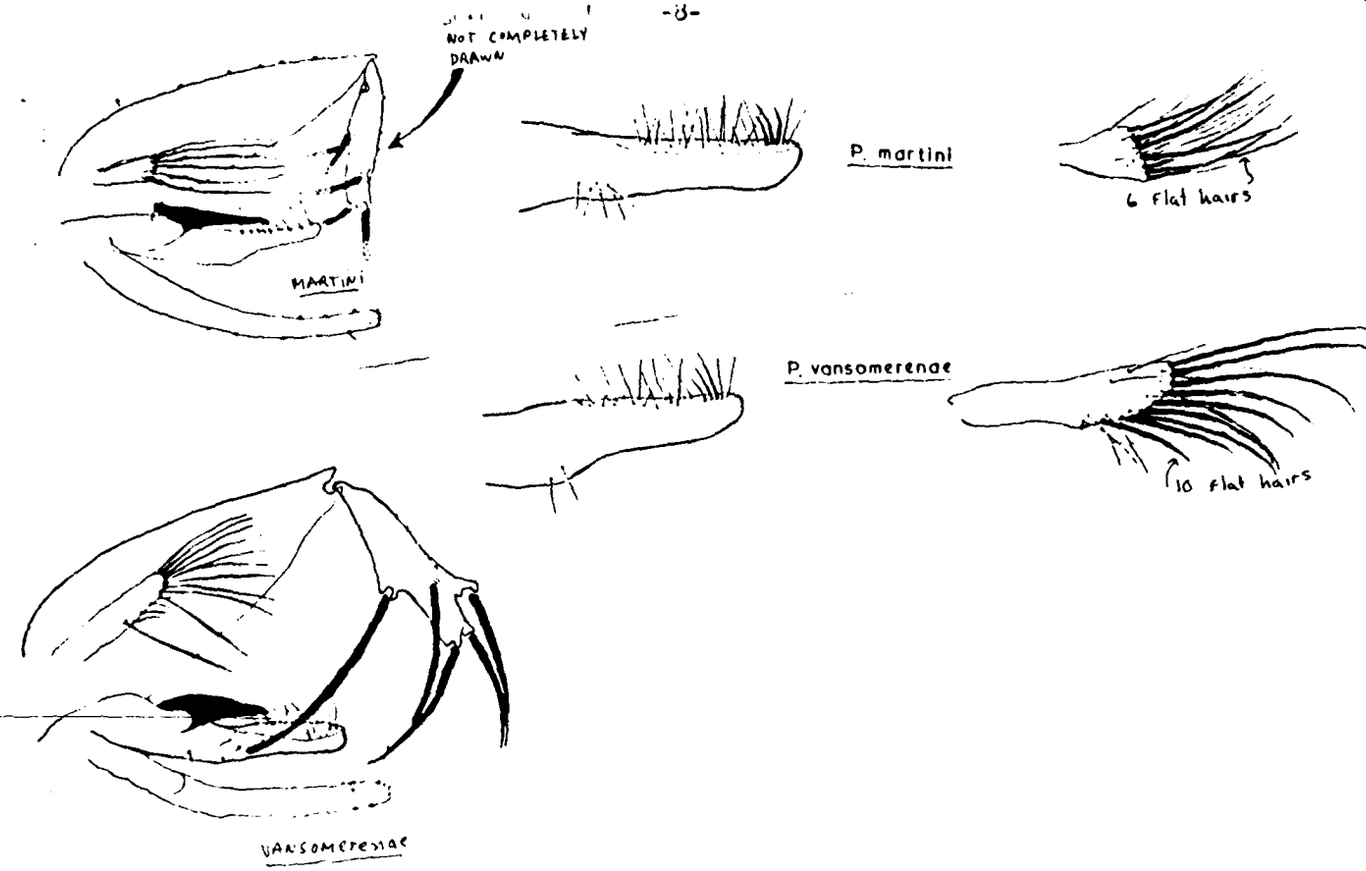
coxite tuft

9. Coxite tuft of 6 flat hairs and about 12 thin ones. . . . . P. martini

Coxite tuft of 10 flat hairs and fewer than 12 thin ones . . . . . P. vansomerenae

* Vector of Kala-azar in Sudan. ALSO CALLED P. ORIENTALIS BY VARIOUS AUTHORS

** Vector of cutaneous Leishmaniasis in Kenya.



10. Paramere forked . . . . . P. rodhaini (FIGURE ON PAGE 1)

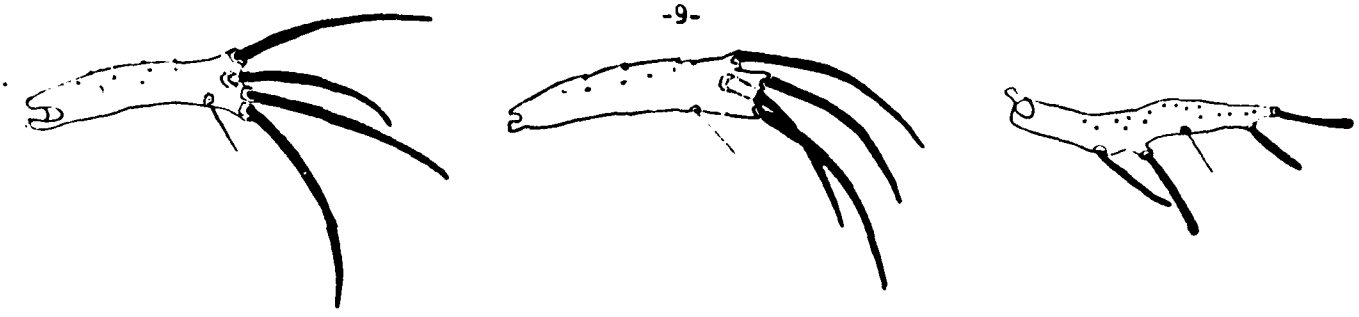
    Paramere simple 11

11. Coxite with persistent setae inserted on a long arm . . . . P. saevus

    Coxite with persistent setae usually absent but, if present, not inserted on an arm or tubercle. (Genus Sergentomyia, in part). 12

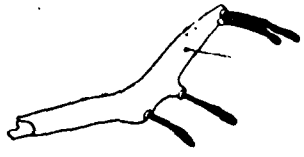
12. Style with 2 terminal spines and 2 median (more or less) major spines . 13

    Style with all major spines well beyond middle of segment 17

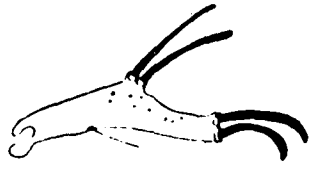


13. Style with 2 median spines well separated 14

    Style with 2 median spines adjacent, close together 15



S. KIRKI

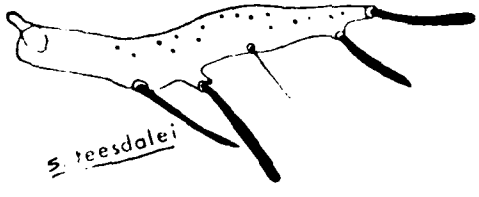


S. WGRAMI

14. Style with 2 terminal spines at end . . . . . S. kirki

    Style with 1 terminal spine at end, the other subterminal . . . . . .

    . . . . . S. teesdalei



S. teesdalei

S. harveyi

15. Style with small seta between the median and terminal groups of major spines . . . . . S. harveyi

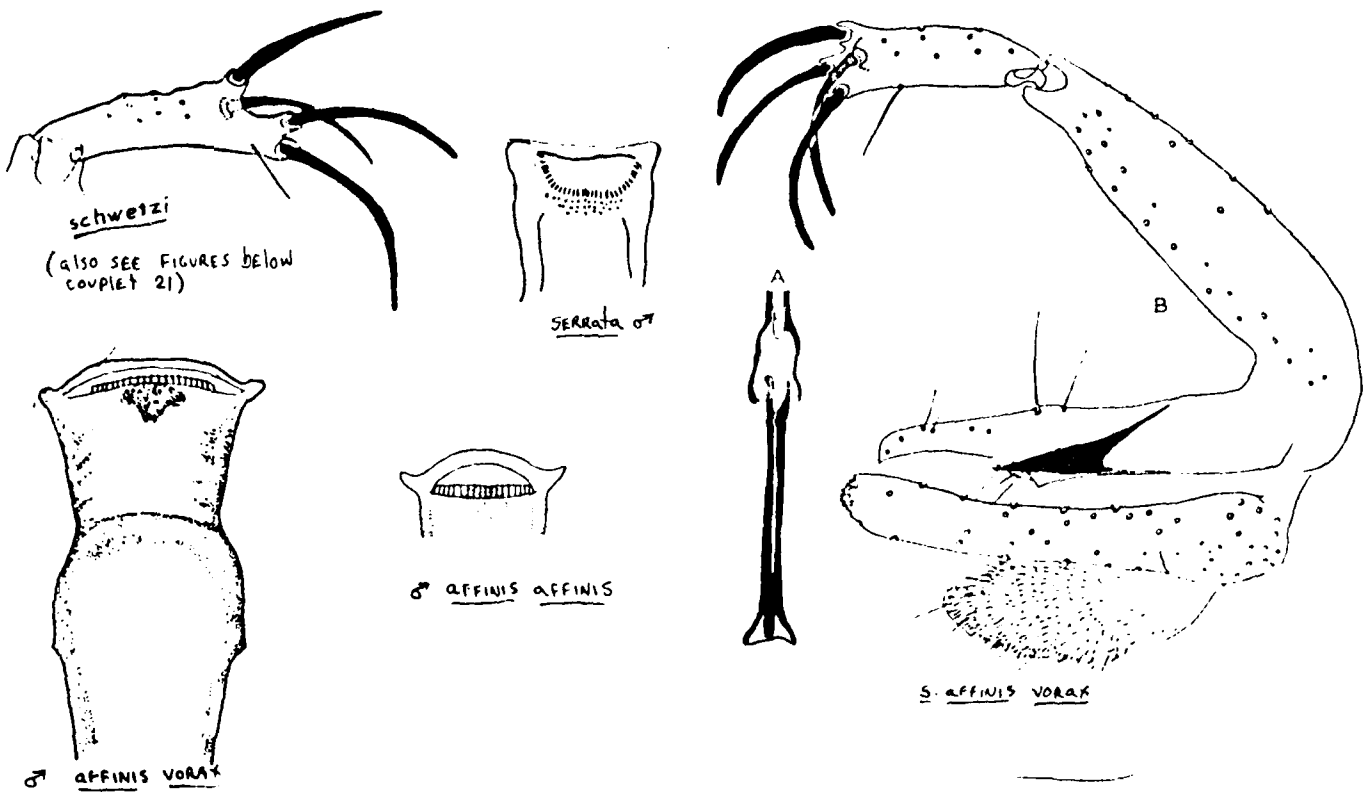
    Style with small seta proximal to, or on same level as, median pair of major spines 16

16. Cibarium with ca. 35 straight pointed teeth arranged in an arc; vertical teeth punctiform in 2-3 rows . . . . . S. serrata

Cibarium with ca. 30 very short teeth; few or no punctiform vertical teeth . . . . . S. ingrami

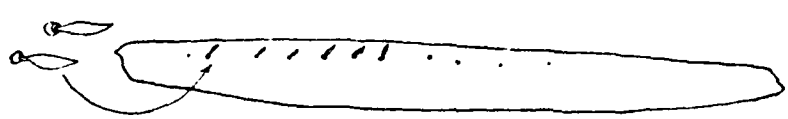
17. Style with 2 terminal spines and 2 others at distal 3/4 . . . . . S. schwetzi (typical form)

Style with all spines terminal or subterminal, i.e. close together. . . 18



18. Front and hind femora armed with short spines . . . . . S. affinis*

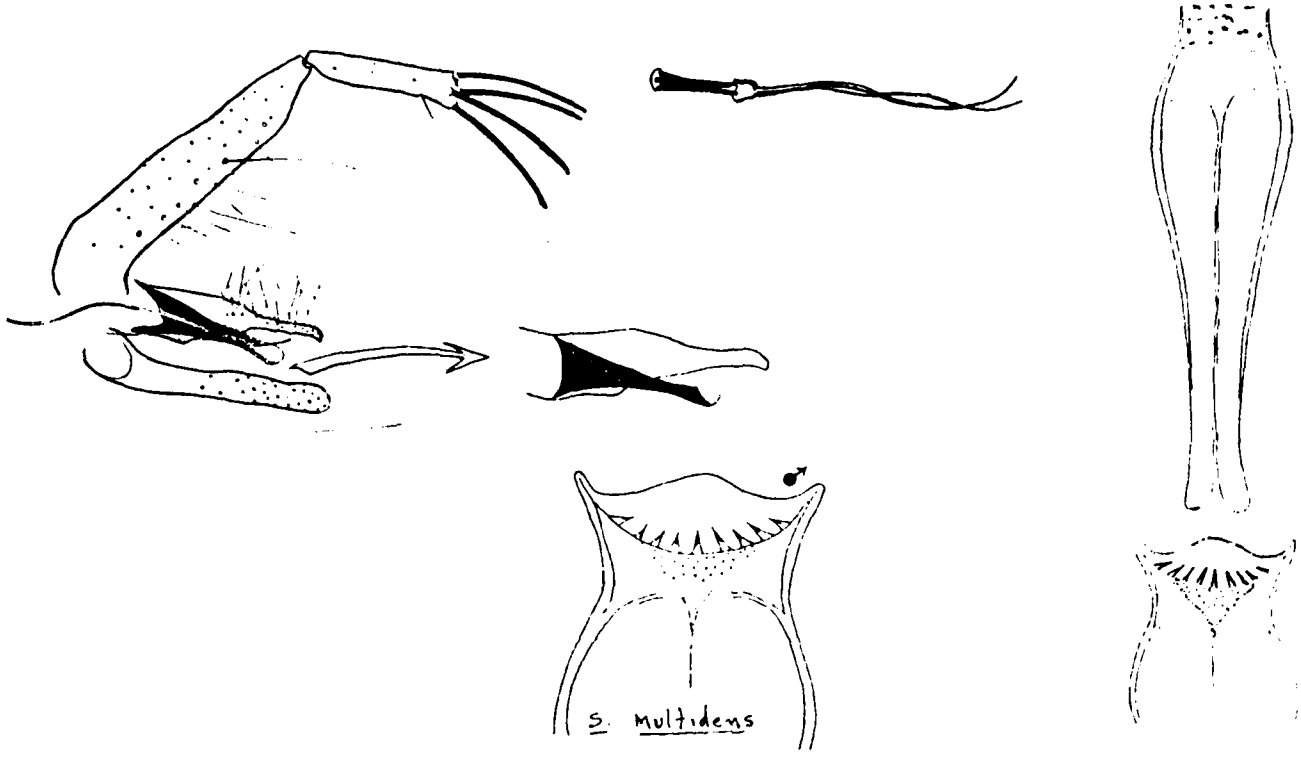
Front and hind femora unarmed 19



* S. affinis affinis and S. affinis vorax occur in Kenya [illegible]

19. Aedeagus with pale tip, rounded, dialated . . . . . S. multidens

Aedeagus with pointed, dark tip 20

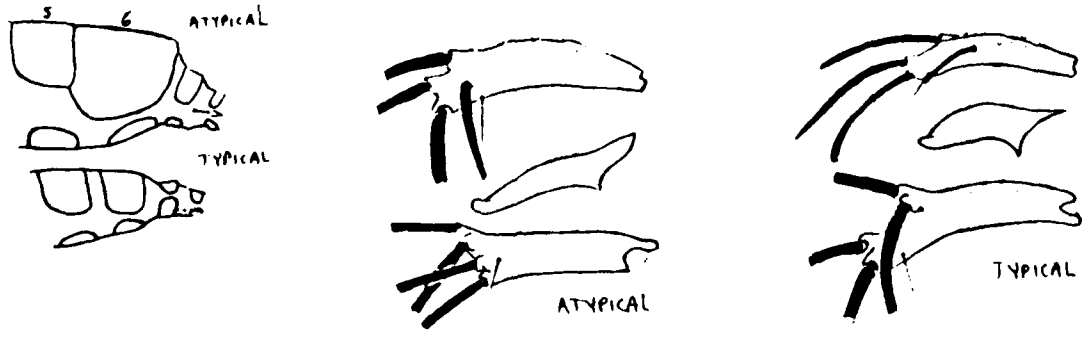


20. Aedeagus finger-shaped 21

Aedeagus conical 24

21. 6th abdominal tergite longer than 5th (atypical form). . . . S. schwetzi*

6 abdominal tergite about same size as 5th 22



22. Paramere (0.16 mm long). About as long as lateral lobe . . . S. yusafi (UNCOMMON IN COASTAL AREAS)

Paramere shorter than lateral lobe 23

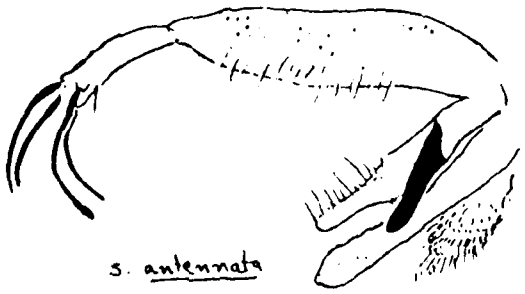
S. yusafi

S. yusafi

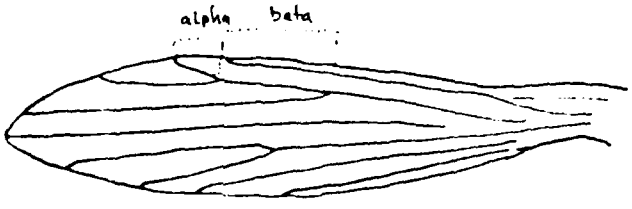
23. Antennal segment 3 short (.095-.100); alar index a/b = 0.50-0.57; pigment patch roughly circular. Palpal segment 4 = to or but little longer than 3 . . . . . S. antennata*

Antennal segment 3 longer (.100-.150); alar index 0.64; pigment patch short and broad. Palpal segment 4 clearly longer than 3. . . . . S. bedfordi

* THERE MAY BE SEVERAL FORMS OF ANTENNATA IN KENYA. TO KEY THEM USE ABONNENC (1972). S. CINCTA IS ONE OF THESE FORMS.



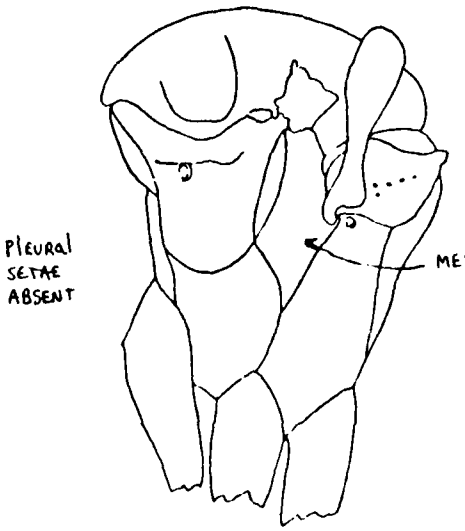
S. antennata



S antennata

24. Pleural setae present* 25

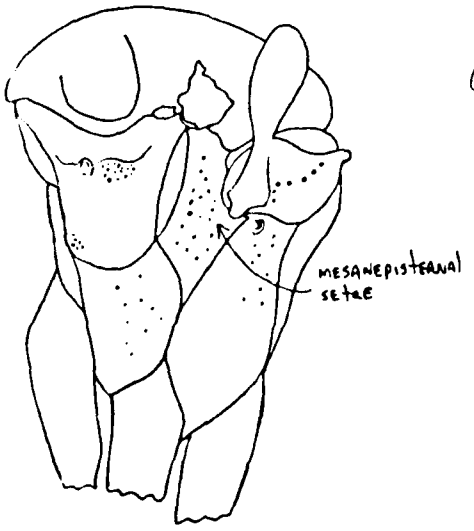
Pleural setae absent 26



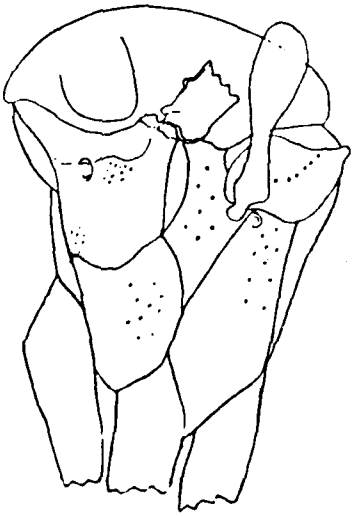
* NOTE. SOME MALES OF SERGENTONYIA AFRICANA magna have pleural setae, but they are never on the mesanepisternum. SEE FIGURE BELOW COUPLET 20 IN FEMALE KEY WHICH SHOWS THE POSITION OF THESE SETAE.

25. Mesanepisternal setae 17-35. Cibarium with 10-15 teeth. Pigment patch small. . . . . S. squamipleuris**

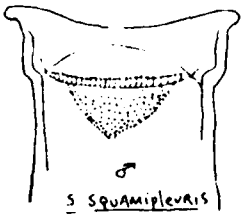
Mesanepisternal setae 1-9. Cibarium with 10-15 teeth. Pigment patch small. . . . . S. inermis



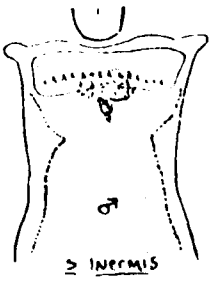
♀ SQUAMIPLEURIS



S. INERMIS



♂ SQUAMIPLEURIS

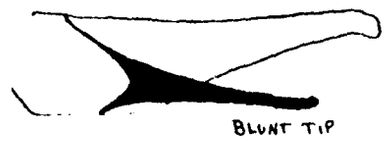


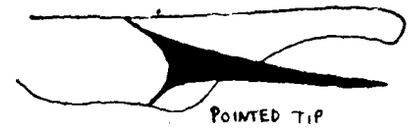
♂ INERMIS

** COMMONLY ATTRACTED TO LIGHT THERE IS AN ATYPICAL FORM WITH ENLARGED 6TH ABDOMINAL TERGITE.

26. Aedeagus with blunt tip 27

Aedeagus with pointed tip 30



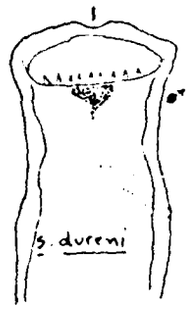


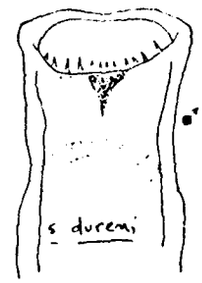
27. Antennal segment III (0.18 - 0.25 mm long), equal to or shorter than labrum 28

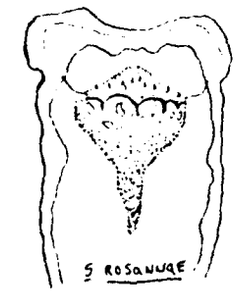
Antennal segment III (0.30 - 0.50 mm long), longer than labrum ... 29

29. Pigment patch well developed with tooth-like thickenings . . . . . *S. rosannae*

Pigment patch invisible or nearly so, no such thickenings . . . . . *S. dureni*

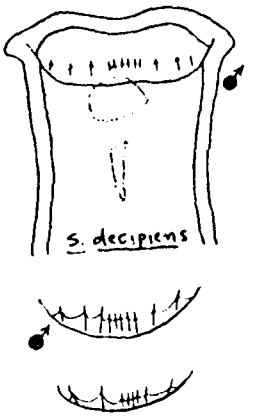


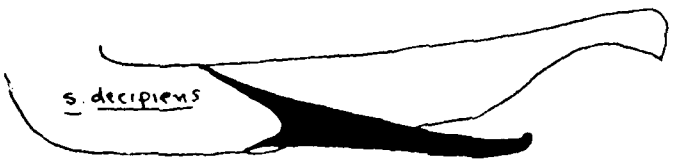




30. Antennal segment III very long (0.40-0.50 long). Cibarium with 15 short needle-like teeth spaced unevenly along a weakly concave posterior arc . . . . . S. garnhami*

Antennal segment III shorter (0.30-0.36 mm long). Cibarium with 4-12 unequal teeth; 3 median ones short and narrow with long and stout lateral teeth . . . . . S. decipiens



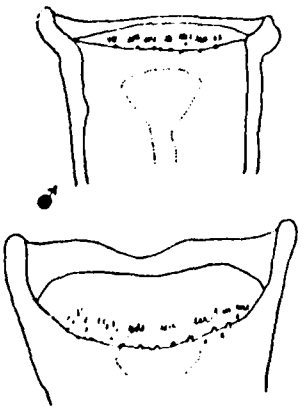


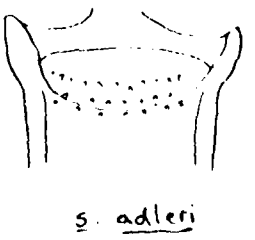
30. Palpal formula 1-2-4-3-5. Antennal segment III shorter or equal to labrum 31

Palpal formula 1-2-3-4-5. Antennal segment III longer than labrum 32

31. Cibraium with 6-7 groups of well-developed denticles. Abdominal tergite 6 considerably larger than 5 . . . . . S. clydei

Cibarium with 10-12 distinct teeth and 2-3 rows of anterior punctiform denticles. Tergites 5 and 6 subequal in size . . . . . S. adleri





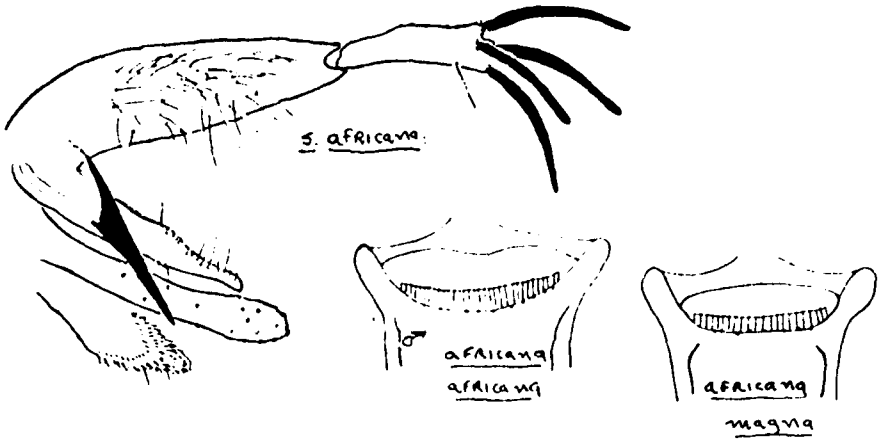
* clydei sometimes bites man, as does garnhami

32. Antennal segment III < 0.28 mm. No pigment plate. Coxite with numerous median and distal setae. Pleural setae present or not .. 33

Antennal segment III > 0.28 mm. Pigment plate present. Coxite with fewer setae. Pleural setae absent 34

33. Pleural setae usually present (see figure below; couplet 20 in female key). Cibarium with 14-23 teeth ..... S. africana magna

Pleural setae absent. Cibarium with 26-36 teeth .... S. africana africana



34. Cibarium with a row of 9 needle-like teeth . . . . . S. meilloni

Cibarium with a row of 17-25 teeth 35

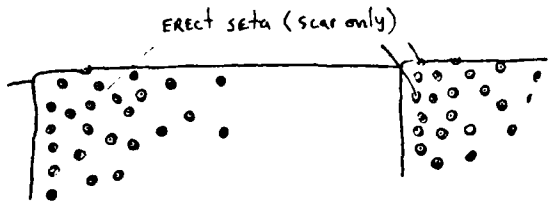
35. Cibarium with a row of 17-20 very short teeth in a palisade; pigment patch short and broad . . . . . S. graingeri

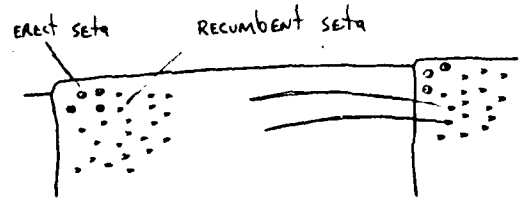
Cibarium with 18-25 fringe-like teeth; pigment patch shaped like a "top" . . . . . S. suberecta

Females*

1. Cibarium without teeth or with only tiny spicules. Pigment patch absent. Hind ends of abdominal tergites 2-6 with erect hairs. Pleural setae absent. (Genus Phlebotomus) 2

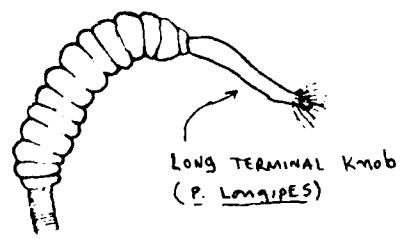
   Cibarium with teeth and pigment patch. Abdominal tergites with recumbent hairs. Pleural setae present or absent. (Genus Sergentomyia) 6

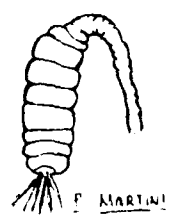




2. Spermathecae with long terminal knob. (Subgenus Larroussius) . . 3

   Spermathecae with short terminal knob 4





3. Wing length 2.8-3.7 mm. Antennal segment 3, 0.34-0.50 mm . . . . . P. aculeatus
   P. guggisbergi
   P. pedifer*
   P. elgonensis

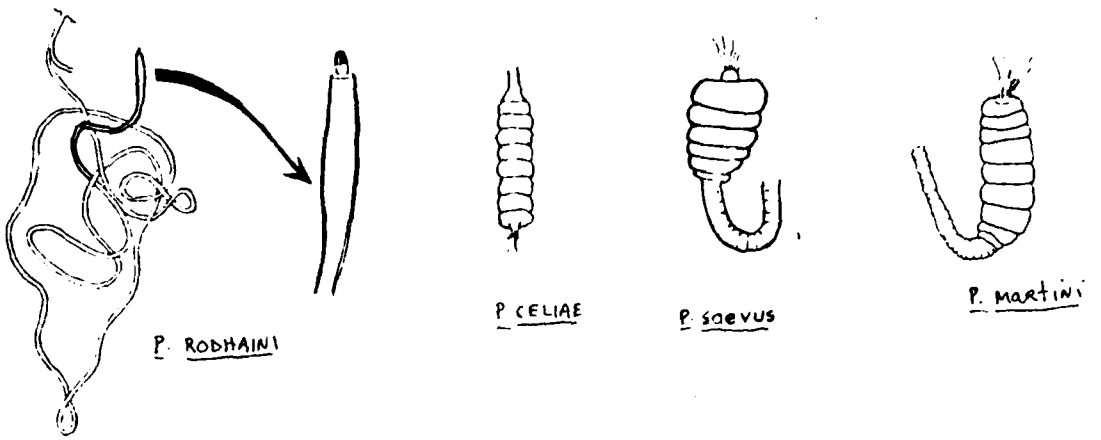
   Wing length 1.9-2.5. Antennal segment 3, 0.25-0.30 mm . . . . . P. langeroni orientalis*

   (= orientalis of various authors)

* FEMALES HAVE BEEN REPORTED BITING MAN

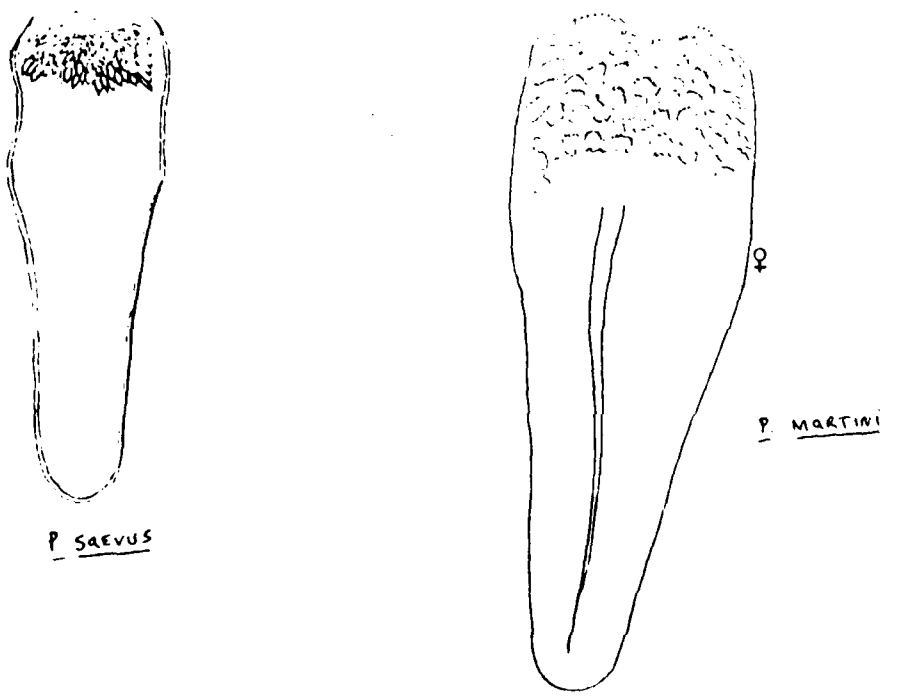
4. Spermathecae smooth-walled, nonsegmented (Subgenus Anaphlebotomus) . . . . . P. rodhaini

Spermathecae with well defined segments 5



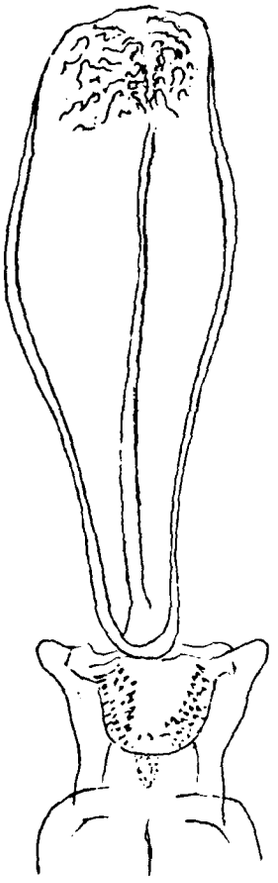
5. Spermathecae with 4-6 segments. Pharynx with armature of coarse teeth. (Subgenus Paraphlebotomus) . . . . . P. saevus

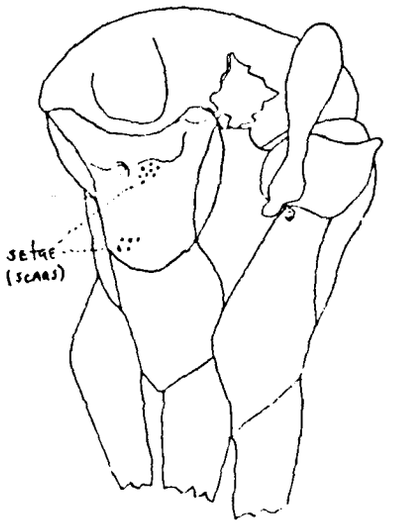
Spermathecae with 9-10 segments. Pharynx with ridges and small teeth. (Subgenus Synphlebotomus) . . . . . P. martini
P. celiae
P. vansomerenae



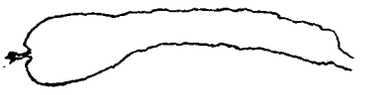
6. Cibarium with lateral longitudinal and irregular rows of teeth. Pleural setae positioned as shown. (Subgenus Parvidens). * . . . . . S. heischi*

Cibarium with a single transerve row of horizontal teeth. Pleural setae usually absent, but if present, differing in number and position from above 7





S. HEISCHI
♀



7. Spermathecae globular with a band of small setae . Pleural setae present. (Subgenus Grassomyia). 8

Spermathecae otherwise, with no setae; pleural setae absent.* . . . 9

GRASSOMYIA - TYPE SPERMATHECA

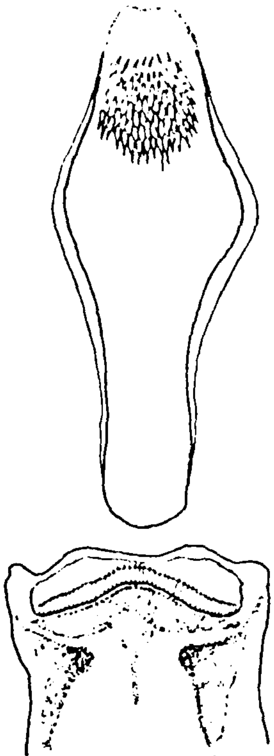
S SQUAMIPLEURIS

* SERGENTOMYIA AFRICANA MAGNA (COUPLET 20) MAY ALSO HAVE ONE GROUP OF PLEURAL SETAE BUT THE SPERMATHECAE ARE QUITE DIFFERENT.

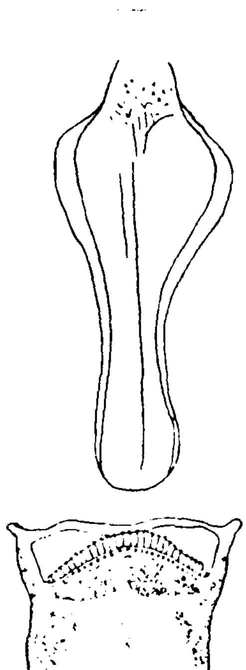
* RARE MAN-BITER

8. Pharynx with numerous stout teeth. Pleural setae on mesanepisterum (SEE ♂ KEY). 17 to 35. Cibarium with 20 to 26 relatively long teeth . . . . *P. squamipleuris*

Pharynx with only a few minute teeth. Pleural setae on mesanepisterum (SEE ♂ KEY) 1-9. Cibarium usually with fewer than 20 horizontal teeth . . . . *P. inermis*



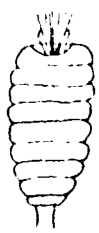
S. squamipleuris



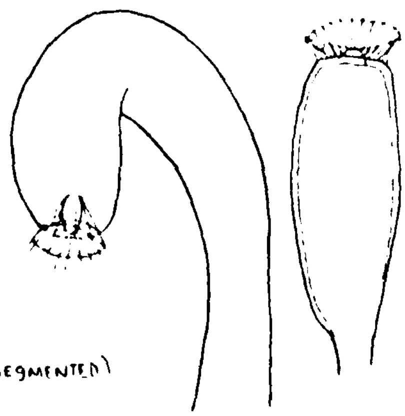
S. inermis

9. Spermathecae clearly segmented 10

Spermathecae smooth walled, nonsegmented 15



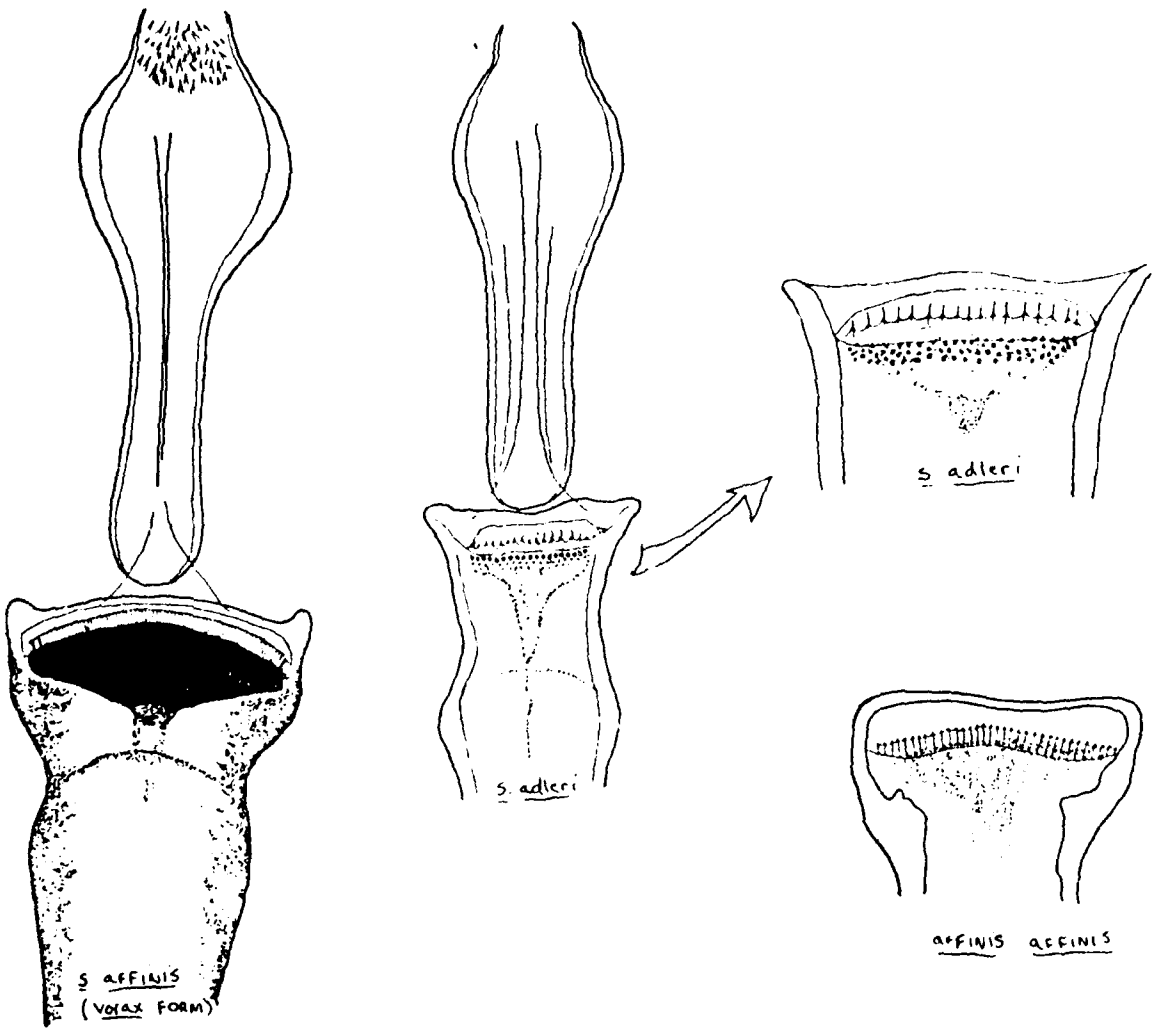
S adleri
(SEGMENTED)



(NONSEGMENTED)

10. Front and hind femora with a row of conspicuous dark spines (see ♂ key). Pharynx with about 50 teeth with long points; cibarium with 34-40 teeth in a comb-like row . . . . . S. affinis*

Front and hind femora unarmed 11



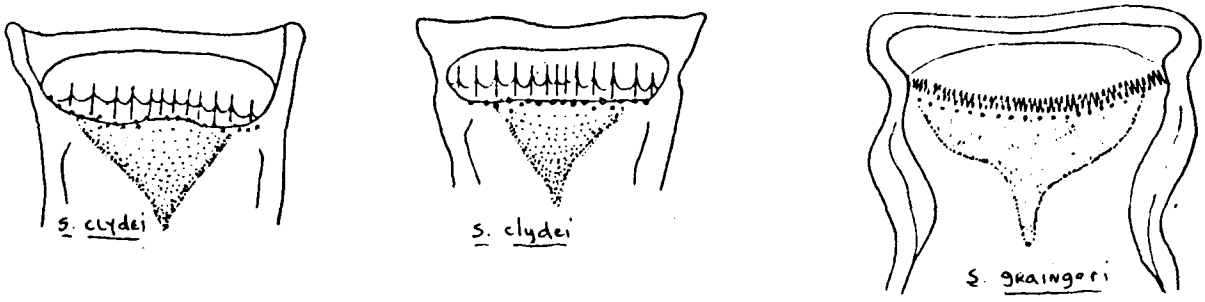
11. Cibarium with 3 or more rows of vertical teeth. . . . P. adleri

Cibarium with 0 to 2 rows of vertical teeth 12

* S. affinis vorax also occurs in Kenya the females of this form have 60 or more horizontal teeth in the cibarium rather than 34 to 40 for S. affinis affinis.

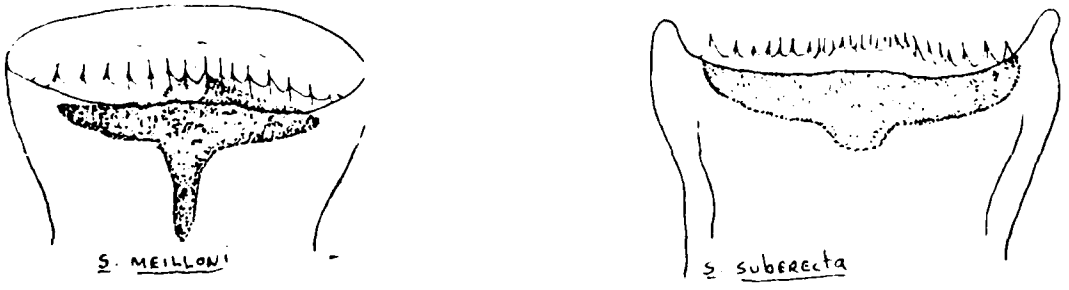
12. Cibarium with well developed row of teeth; no vertical teeth 13

Cibarium with vertical teeth 14



13. Cibarium with 14-16 stout, pointed teeth; pigment patch large and mushroom shaped . . . . . S. meilloni

Cibarium with 25-27 teeth, the lateral ones larger and farther apart than the median teeth . . . . . S. suberecta

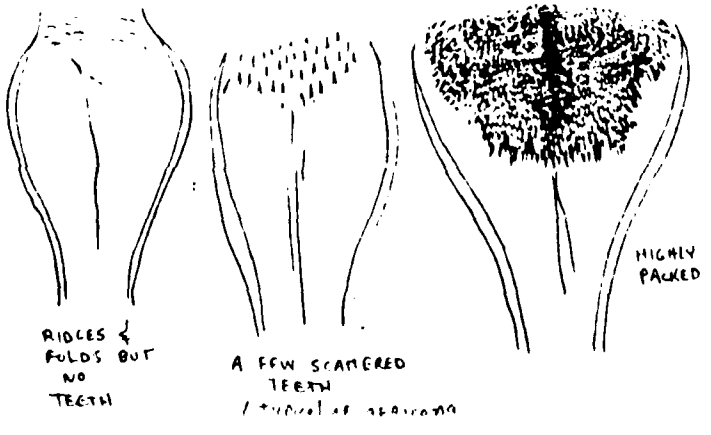


14. Cibarium with 25 or more horizontal teeth. Pigment patch in the form of a pointed helmet . . . . . S. graingeri

Cibarium with 20 or fewer horizontal teeth. . . . . S. clydei

15. Pharynx armed with numerous teeth in highly packed rows. 16

Pharynx unarmed or with only a few scattered distinct teeth . . . 19

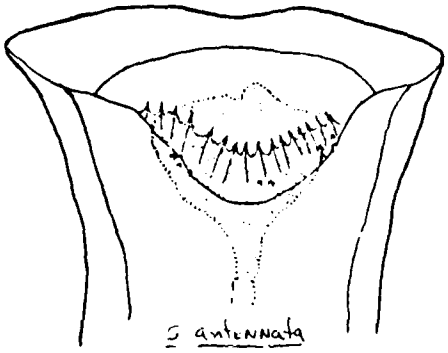


16. Cibarial teeth suboval, monomorphic 17

Cibarial teeth in middle smaller than lateral teeth 18
(N.b. S. antennata and bedfordi common; S. yusafi and gracilis rare)

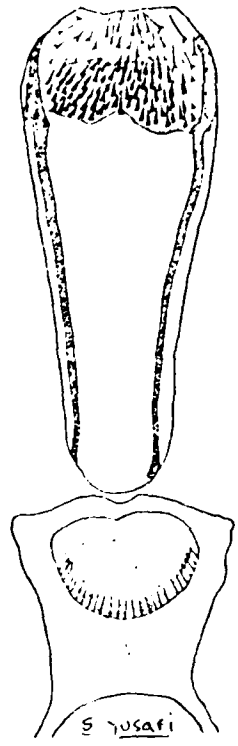
17. Antennal segment III < .100 mm; palpal segment 4 somewhat > or = 3 . . . . . S. antennata*

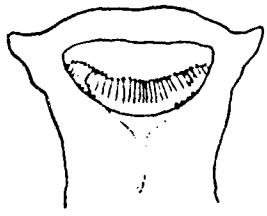
Antennal segment III > .100 mm (coastal sp.) . S. yusafi (rare)



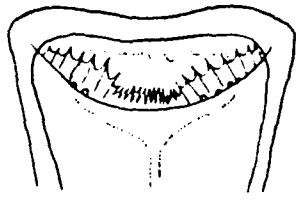
S antennata (cincta form)

S. antennata





S. antennata antennata



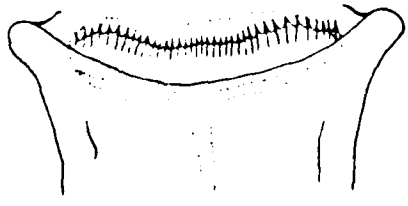
S. bedfordi bedfordi

S yusafi

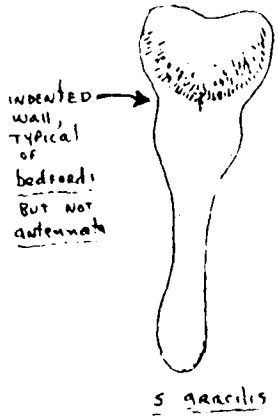
18. Cibarium with 26-28 small pointed teeth, the median ones smaller than laterals . . . . . S. gracilis (rare)

Cibarium with 24-28 teeth on an arc, the 12-13 median teeth small, the lateral teeth stout, palpal segment 4 clearly > 3 . . . . . S. bedfordi*

* S. bedfordi w/ indented pharyngeal wall (compare Quate, Fig. 19 B & 20C)



S bedfordi congolensis, most ♀♀ in Kenya correspond to the congolensis form. According to Minter, congolensis



S gracilis

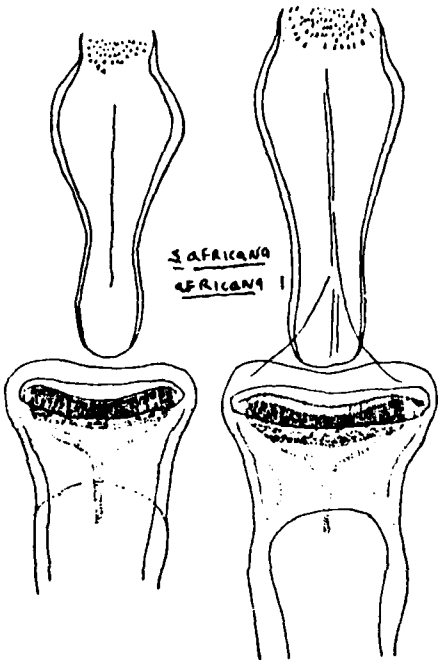
S gracilis

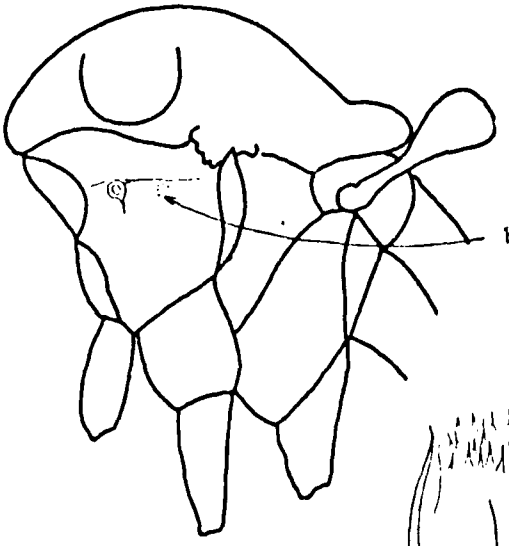
19. Pharynx armed with a small number of scattered denticles 20

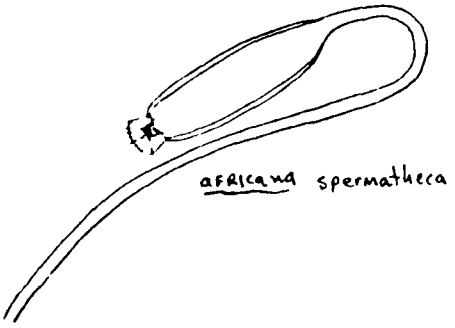
    Pharynx unarmed or with fine folds, ridges, or broad scales having finely denticulate (dot-like) posterior borders 21

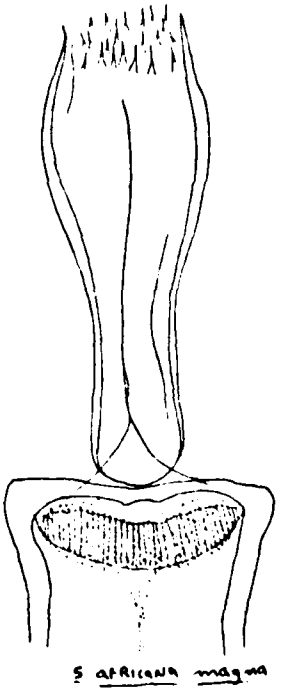
20. Cibarium armed with about 60 teeth; pigment plate black, often obscuring teeth ..... S. africana africana

    Cibarium armed with not more than 45 teeth. Pigment patch usually brown; 1-6 insertions (ie. setal scars) on thoracic pleurae .... S. africana magna*









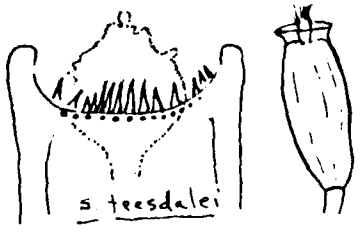
* Bonnenc (1972) elevates this subspecies to species rank, calling it S. magna. Not all specimens have pleural setae in Ethiopia. Specimens in Kenya may or may not show this variation. It is not clear whether one or both of the forms in couplet 20 occurs in Kenya.

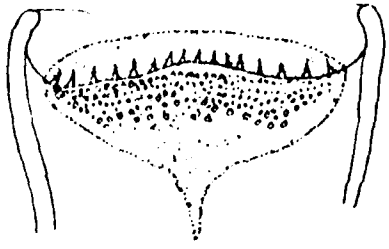
21. Antennal segment III very long (.390-.44 mm). 22

Antennal segment III long (.09-.38 mm) 23

22. Cibarium with 11-12 teeth and only 1 row of anterior vertical teeth . . . . . S. teesdalei

Cibarium armed with 16 strong teeth and w/ 6-8 rows of anterior vertical teeth. . . . . S. multidens





23. Pigmented plate well developed, filling the entire width of the cibarium 24

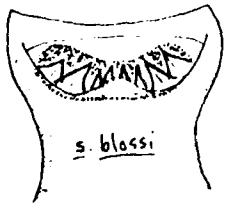
Pigmented plate narrow or absent 29

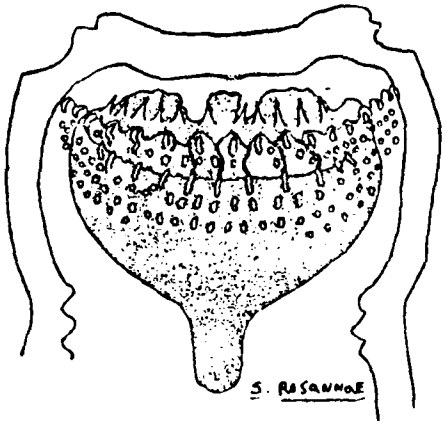
24. Cibarium with 8 to 18 teeth 25

Cibarium with 20 teeth. 28

25. 8 cibarial teeth, the 4 median ones short and narrow, the lateral teeth stout and wide pigmented plate bilobed. . S. blossi

10-15 cibarial teeth; pigment plate not bilobed 26



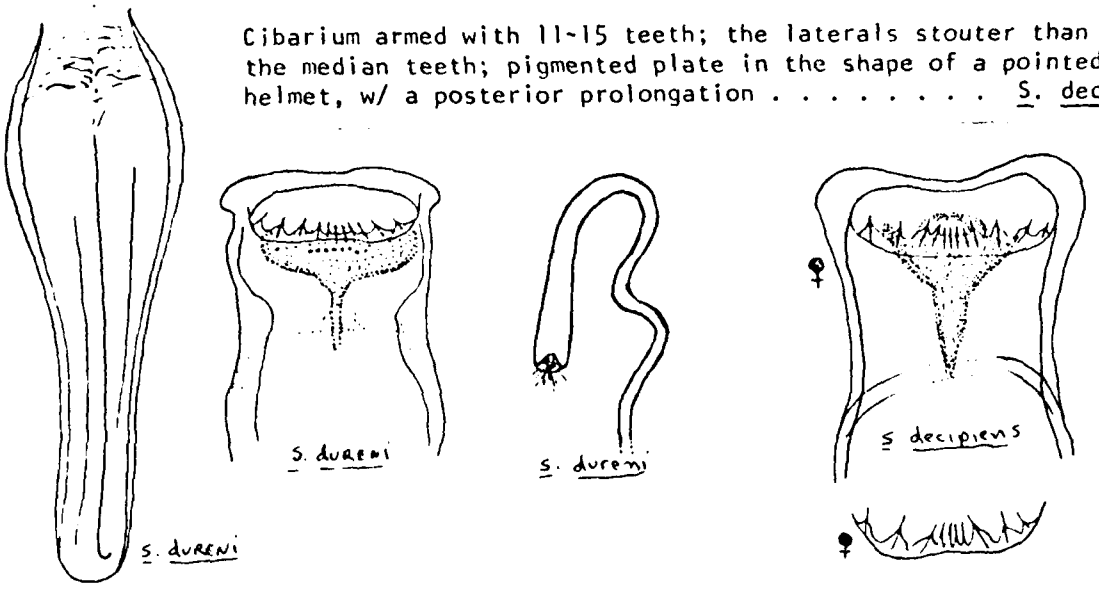


26. Cibarium armed w/ 10-12 short retractile teeth, scarcely visible; very stout, rounded tooth-like folds present anterior to the cibarial teeth; 3-6 rows of anterior vertical teeth. Pigmented plate large and massive . . . . . S. rosannae

No rounded tooth-like folds in the cibarium 27

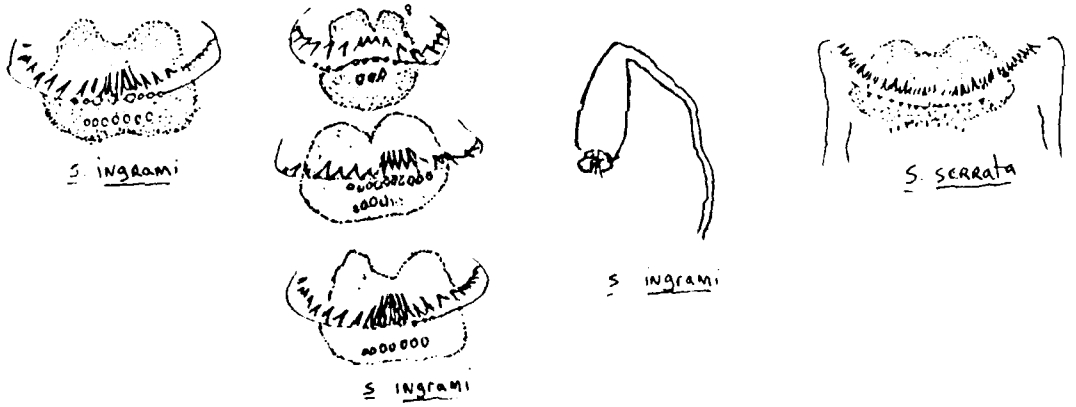
27. Cibarium armed w/ 12-14 broad, needle-like teeth and 1 or 2 rows of anterior vertical teeth; pigmented plate a narrow transverse band . . . . . S. dureni

Cibarium armed with 11-15 teeth; the laterals stouter than the median teeth; pigmented plate in the shape of a pointed helmet, w/ a posterior prolongation . . . . . S. decipiens



28. Cibarium armed with 24-28 polymorphic teeth, 2 or 3 rows of anterior vertical teeth, of which the central teeth are noticeably larger; a few erect hairs on abd. tergites 2-6 . . . . . S. ingrami

Cibarium with 40-52 teeth and 3 rows of subequal vertical teeth. No erect hairs on abdominal tergites 2-6. . . S. serrata

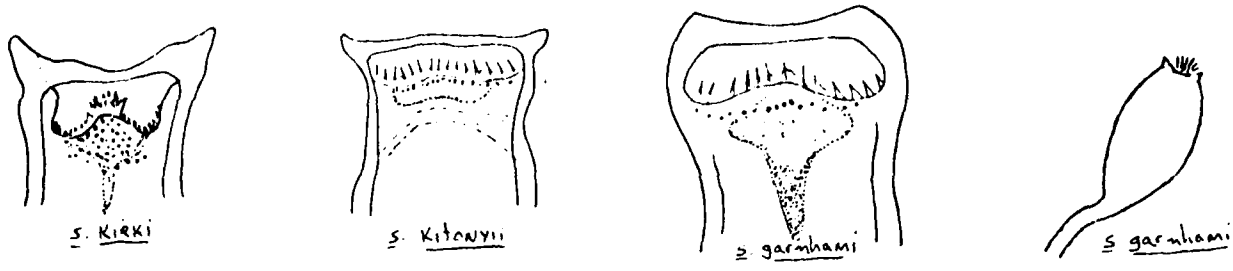


29. Cibarium with 1 or several rows of vertical teeth, or an unorganized group of teeth 30

Cibarium without vertical teeth 32

30. Cibarium with 22-24 polymorphic teeth on an M-shaped sinuous line; numerous vertical teeth not arranged in rows . . . . . S. kirki

Cibarium with a transverse row of vertical teeth. 31

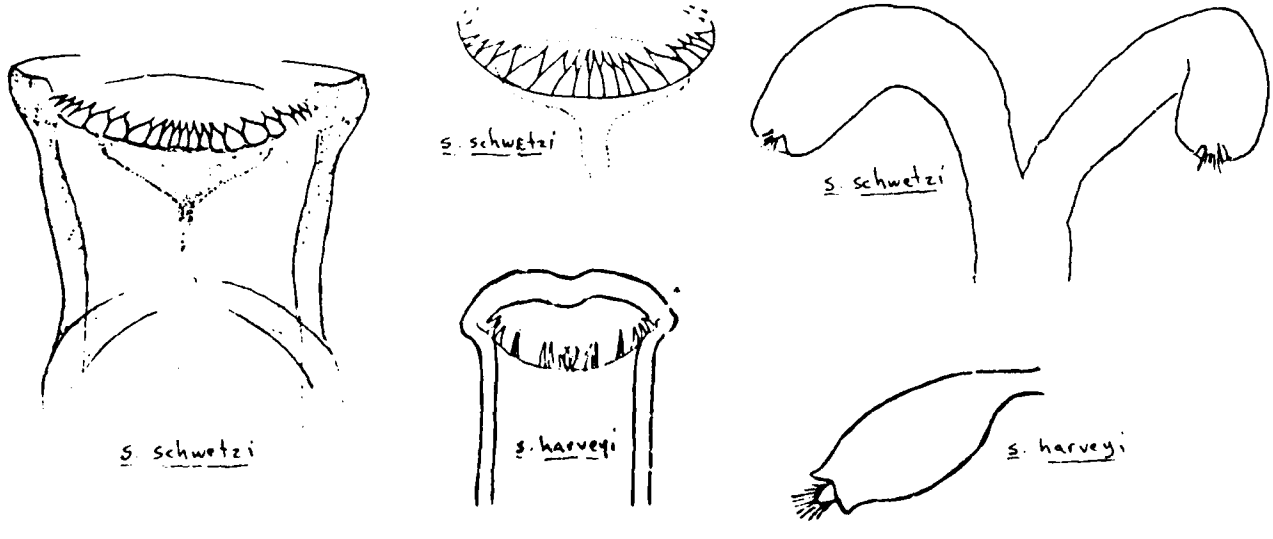


31. Pigment patch heart-shaped; antennal segment 3 0.26 - 0.32 mm long; cibarium with 15 long subequal needle-like teeth on a slightly sigmoidal. A row of anterior vertical teeth. . . . . S. kitonyii

Pigment patch, top-shaped; cibarium 13 stout pointed teeth on a slightly sinuous line and a row of 14-16 vertical teeth . . . . . S. garnhami (A RARE MAN-biter)

32. Spermatheca tubular with the terminal knob embedded in the body. Cibarium with 16-18 teeth. . . . . S. schwetzi*

Spermatheca cylindrical. Cibarium with 11 teeth in an arc . . . . . S. harveyi



* SOMETIMES bites MAN. THE ATYPICAL AND TYPICAL FORMS ARE separated only in the

Appendix II

Sand Flies of the Central Amazon of Brazil. 2. Description of Lutzomyia (Trichophoromyia) ruii n. sp. (Diptera: Psychodidae)[1]

Jorge R. Arias[2]

David G. Young[3]

Abstract

Lutzomyia (Trichophoromyia) ruii Arias & Young n. sp. is described and illustrated from both sexes commonly found near Manaus, Brazil. Females are not anthropophilic. Information on seasonal distribution is given. Lutzomyia (Trichophoromyia) melloi (Causey & Damasceno) is a junior synonym of Lutzomyia (T.) inini (Floch & Abonnenc) (NEW SYNONYM).

Introduction

Sherlock and Guitton (1970) reviewed the phlebotomine subgenus Trichophoromyia Barretto, 1962, described a new species, and provided a key to the males of 18 species. Two of them, Lutzomyia (T.) melloi (Causey & Damasceno, 1945) and Lutzomyia (T.) inini (Floch & Abonnenc, 1943) were separated in couplet 4 by genitalic characters. We now believe, however, that L. inini is a senior synonym of L. melloi based on recent determinations of males from Brazil and French Guiana by us and M. Emile Abonnenc (NEW SYNONYM). Recently, Young (1979) described 3 new Trichophoromyia spp. from southern Colombia. Here, we describe another species which is rather common in the vicinity of Manaus, Brazil.

Lutzomyia ruii is named in honor of Mr. Rui Alves de Freitas of the Instituto Nacional de Pesquisas da Amazônia for his dedicated service in the continuing study of phlebotomines and leishmaniasis in the Amazon Basin of Brazil.

Terminology follows that of Young (1979). The description is based on the holotype and allotype with ranges of some measurements of paratypes (111♂♂, 11♀♀) given in parentheses. Measurements are in millimeters.

Lutzomyia (Trichophoromyia) ruii Arias & Young, n. sp.

Fig. 1-9

Male: A large dark sand fly; pleura moderately pigmented, mesonotum strongly infuscated. Wing length 2.10 (2.06-2.18); width 0.56 (0.53-0.56). Cibarium with about 12, scattered dot-like remnants of vertical teeth; cibarial arch distinct only at sides, without pigment patch. Pharynx 0.18, unarmed. Head height ; width . Eyes large, separated by 0.126 or by distance equal to 7.1 facet diameters. Flagellomere I, 0.23 (0.21-0.24), combined length of II + III slightly longer than I; ascoids with very short distinct posterior spurs, the distal tips of ascoids on II reaching beyond flagellomere, on all flagellomeres except last 3. Labrum 0.23 (0.21-0.24) long. Length of palpal segments (holotype): 1 (0.04), 2 (0.10), 3 (0.40), 4 (0.06), 5 (0.14); palpal sensilla at end of segment 2 and along middle third of 3. Pleura with 15 (7-15) upper and 2 (2-4) lower episternal setae. Length of wing vein sections: alpha 0.56 (0.49-0.58), beta 0.28 (0.26-0.33), delta 0.42 (0.28-0.42), gamma 0.22 (0.22-0.28). Length of femora, tibiae and basitarsi of holotype: foreleg, 0.86, 1.13, 0.68; midleg, 0.79, 1.35, 0.83; hind leg, 0.86, 1.58, 0.90. Genitalia: style 0.23 long, with 4 major spines arranged as shown, no subterminal seta. Coxite ca. 0.39 long x 0.12 wide, bearing a median group of ca. 45 long slender hairs. Paramere simple as shown. Aedeagus broad, well pigmented. Genital pump 0.18 (0.15-0.18)

long, each filament 0.87 (0.87-1.02) long or 4.8 x length of pump, tip simple. Lateral lobe 0.38 (0.38-0.40) long. Cercus as shown.

Female: Wing length 2.20 (2.20-2.40); width 0.64 (0.64-0.75). Cibarium, with 10-12 slender pointed, more or less equidistant horizontal teeth and 20-30 vertical teeth, median teeth larger than others; cibarial arch complete; pigment patch well infuscated. Pharynx 0.19 (0.18-0.20) long, unarmed. Head height ; width . Eyes large, separated by 0.55 or by distance equal to 7.2 facet diameters. Flagellomere I 0.20 (0.20-0.25) long, II + III slightly longer than I; ascoids as in male, on all flagellomeres except last 3. Labrum length 0.34 (0.34-0.43). Length of palpal segments of allotype: 1 (0.05), 2 (0.15), 3 (0.19), 4 (0.06), 5 (0.14); palpal sensilla as for male. Pleura with 9 (9-17) upper and 2 (2-4) lower episternal setae. Length of wing vein sections: alpha 0.70 (0.64-0.78), beta 0.27 (0.27-0.30), delta 0.49 (0.41-0.55), gamma 0.25 (0.20-0.29). Length of femora, tibiae and basitarsi of allotype: foreleg, 0.75, 1.01, 0.71; midleg, 0.75, 1.24, 0.83; hindleg, 0.83, 1.46, 0.94. Spermathecae annulated except for smooth subterminal portion; individual ducts very long, ca. 6.5 x length of spermatheca; common duct shorter than spermatheca.

Type Data: Holotype male (DCDC-419, no. 1).

Reserva Ducke, 26 km E of Manaus, Amazonas, Brazil; light trap; 6 Dec. 1977; J. Arias, R. Freitas & J. Vidal colls. Allotype female (Ducke-CDC, no. 2). Same data but taken on 18 June 1976. Paratypes (12♂♂, 30♀♀). Reserva Ducke, Reserva Campinas (43 km NE of Manaus), at km 30 and 4 km S of km 56 Rodovia Torquato Tapajós-

Estrada AM-010; CDC light traps, flight traps, soil emergence traps, and animal burrows; 1974 to 1979, J. Arias et al. colls.

Holotype, allotype and 20 paratypes to be deposited at Instituto Nacional de Pesquisas da Amazonia, Manaus. Other paratypes in Brazilian collections at Universidad Federal de Minas Gerais and São Paulo; Instituto Oswaldo Cruz, Rio de Janeiro; Instituto Evandro Chagas, Belém; Museu Paraense Emilio Goeldi, Belém. Remaining paratypes in collections of U.S. National Museum (Natural History), Washington; Florida State Collection of Arthropods, Gainesville, Florida; Instituto de Salude Publica, Lima; British Museum (Natural History), London. All specimens slide mounted.

Discussion:

We associated the sexes of L. ruii on the basis of collecting data. No other Trichophoromyia spp. were taken at the study site 30 km NE of Manaus during a 62 week period of time.

This species occurs with Lutzomyia (Trichophoromyia) ubiquitalis at Reserva Campinas. The males differ readily by the shape of the parameres (more slender in ubiquitalis) and by the length of the genital filaments (shorter than 4X length of pump for ubiquitalis; over 4.5X for ruii).

The females of ruii and ubiquitalis closely resemble each other in nonsexual traits of color, cibarial armature and wing venation (This generally is true for the other Trichophoromyia females as well). However, the shorter sperm ducts and larger terminal knob of the spermathecae of ubiquitalis (Fig. 10) easily serve to distinguish the females. Floch and Abonnenc (1943, 1952)

describe and illustrate the ubiquitalis female as Phlebotomus cauchensis, an accepted junior synonym.

From the Trichophoromyia males having a median tuft of numerous coxite setae such as L. inini, L. auraensis (Mangabeira, 1942) and L. loretonensis (Llanos, 1964), L. ruii differs in the shape of the parameres (see Sherlock & Guitton, 1970, for figures of these other males).

Light trap catches from the site, 30 km NE of Manaus, during a 62 week period (Fig. 11) indicated population peaks of L. ruii adults from August-November, 1977 and February-March and August, 1978. Females do not attack man and their preferred hosts remain unknown.

We wish to thank M. Emile Abonnenc for identifying a male of L. inini collected in French Guiana by Dr. T.H.G. Aitken. Also, we appreciate the valuable assistance of Mr. João Ferreira Vidal in the field and laboratory.

Literature Cited

Floch, H. & E. Abonnenc. 1943. *Phlébotomes de la Guyane Française* - v. Inst. Pasteur Guyane Pub. No. 61. 30p.

_____. 1952. Dipteres phlébotomes de la Guyane et des Antilles Françaises. Faune de L'Union Français, no. 14. Office de la Recherche Scientifique Outre-Mer. 207p.

Sherlock, I.A. & N. Guitton. 1970. Notas sôbre o subgênero Trichophoromyia Barretto, 1961 (Diptera, Psychodidae, Phlebotominae). Rev. Brasil. Biol. 30:137-150.

Young, D.G. 1979. A review of the bloodsucking psychodid flies of Colombia (Diptera: Phlebotominae and Sycoracinae). Univ. of Florida Agr. Exp. Sta. Tech. Bull. 806. 266p.

Footnotes

1) Research was partly funded by CNPq Grant SIP 08/131; INPA Project 2017/103, and U.S. Army Research and Development Contract DADA 17-72-C-2139.

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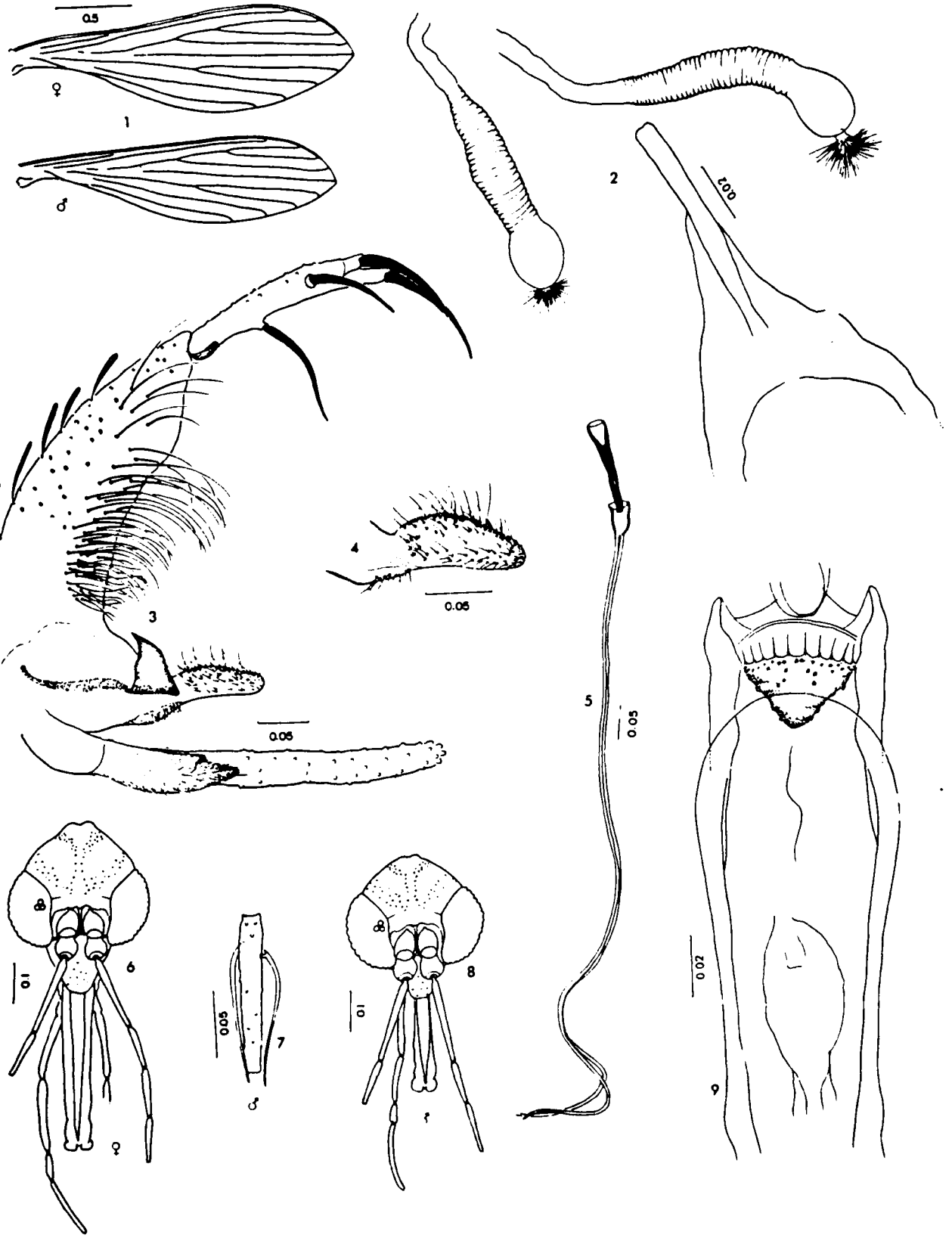
3. Dept. of Entomology and Nematology, University of Florida, Gainesville, Florida U.S.A. 32611.

Figures

Fig. 1-9. Lutzomyia ruii Arias & Young n. sp. 1) Female and male wing. 2) Spermathecae and genital fork, sperm ducts not visible. 3) Male genitalia, lateral view. 4) Paramere, a different, larger view than shown in Fig. 3. 5) Genital pump and filaments. 6) Female head. 7) Male flagellomere II showing ascoids. 8) Male head. 9) Female cibarium. All figures drawn from specimens found at Reserva Ducke. Scale in mm.

Fig. 10. Spermatheca of Lutzomyia ubiquitalis from Belém, Pará, Brazil. The other spermatheca was not drawn. Scale in mm.

Fig. 11. Seasonal distribution of Lutzomyia ruii n. sp. based on light trap catches at 1 m and 15 m heights above ground at Reserva Ducke (1977-1978).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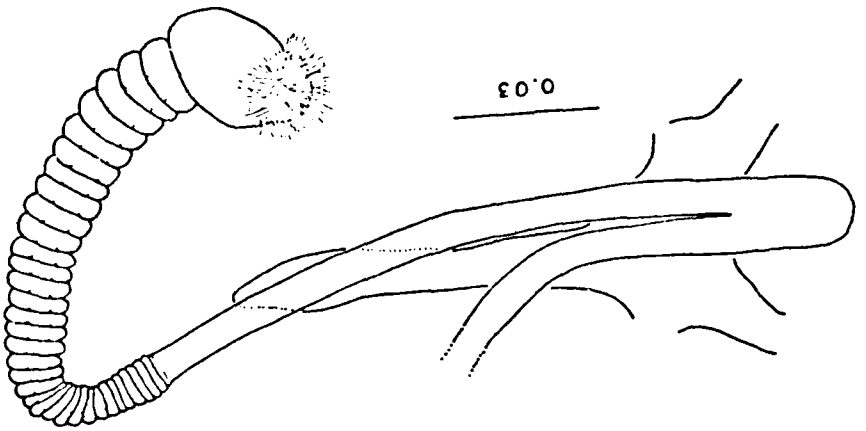

Fig. 10

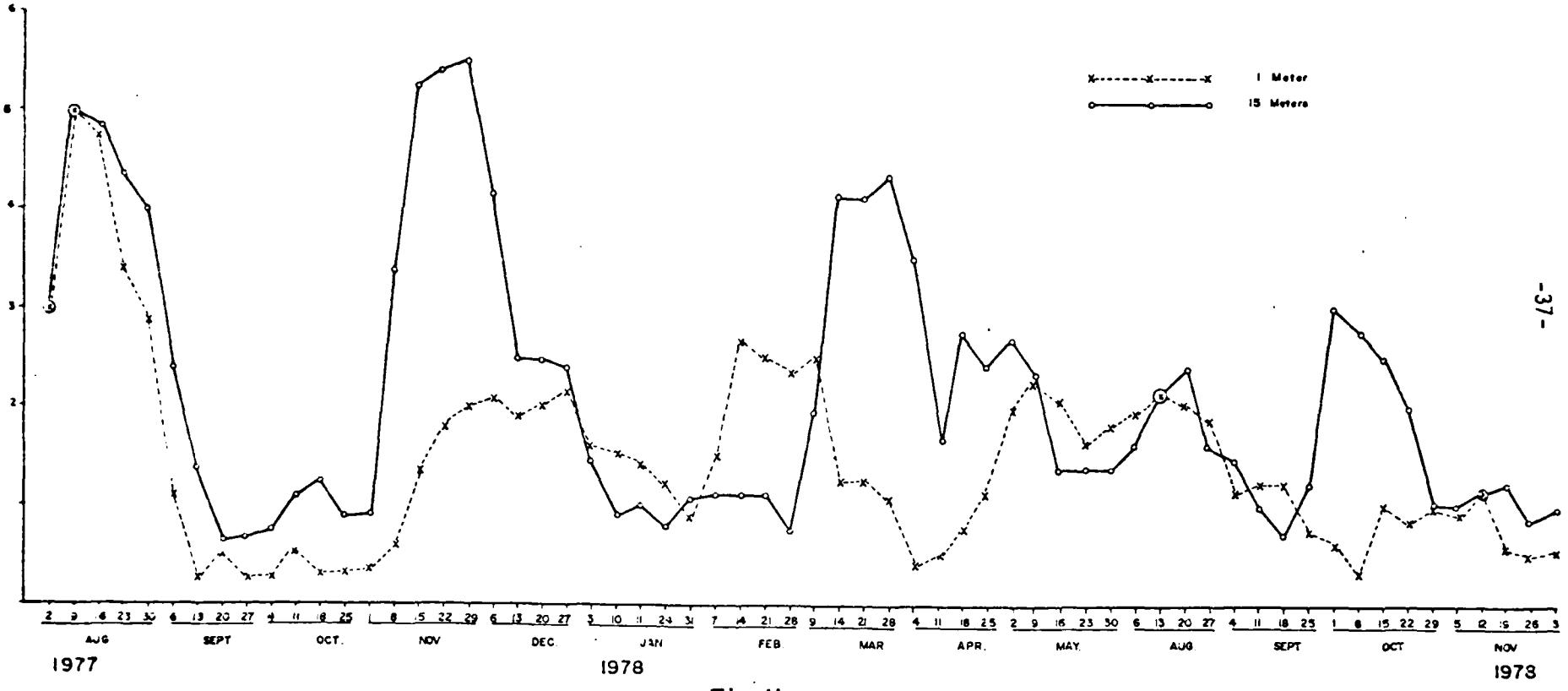
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Fig. 11

Appendix III

A NEW PHLEBOTOMINE SAND FLY IN THE *Lutzomyia flaviscutellata* COMPLEX FROM NORTHERN BRAZIL (DIPTERA: PSYCHODIDAE)[1]

D.G. Young[2]

J.R. Arias[3]

Sand flies of the *Lutzomyia (Nyssomyia) flaviscutellata* complex are important as vectors of *Leishmania mexicana* to small mammals and man in the neotropics. Lainson and Shaw (1968) and Ward et al. (1973, 1977) incriminated *Lu. flaviscutellata* (Mangabeira) as the vector of *Leishmania mexicana amazonensis* in Brazil. Tikasingh (1975) recovered this parasite, or a similar one, from wild caught *flaviscutellata* females in Trinidad. *Lu. olmeca olmeca* (Vargas and Nájera) in Mexico and Central America and *Lu. o. bicolor* Fairchild and Theodor in Panama are proven or suspected vectors of *Leishmania mexicana mexicana* and *L. m. aristedesi*, respectively. Lainson and Shaw (1979) reviewed the epidemiology of these diseases, noting that "it appears that subspecies of *L. mexicana* are principally parasites of sylvatic rodents . . . transmission occurs at ground level, where man also becomes infected."

Aitken et al. (1975) isolated Pacui virus and several other arboviruses from wild caught *Lu. flaviscutellata* females near Belém, Brazil. The Disney trap, an animal-baited oil trap routinely used by them and other workers, is excellent for capturing sand flies in this complex (Disney, 1966).

Fairchild and Theodor (1971) and Lewis (1975) provided information on the taxonomy and distribution of these forest sand flies, the latter author observing among other things, "the presence of many erect hairs

on abdominal tergite 2, while there are none on segment 5. These features and those already known make this complex a remarkably distinct one." The striking coloration of these sand flies, i.e. the pale pleura, coxae and posterior mesonotum contrasting with very dark head, abdomen, and legs, often allows workers to identify specimens without the aid of magnification.

The form described here as a subspecies of *Lu. olmeca* occurs with *Lu. flaviscutellata* at Manaus, Brazil, and vicinity. All measurements are given in millimeters.

Lutzomyia olmeca nociva Young and Arias n.ssp.

Fig. 1-10

Male holotype: Wing length 1.89; width 0.55. Except for pale coxae, pleura, posterior part of mesonotum, rest of insect dark. Head height from vertex to tip of clypeus, 0.36; width 0.33. Eyes large, separated by only 0.04 or by distance = to 2 facet diameters. Interocular suture absent. Flagellomere I, 0.31 long, combined length of II + III = 0.29; ascoids simple, tips of those on II ending before end of flagellomere, on all flagellomeres except last 3. Labrum 0.16 long. Length of palpal segments: 1 (0.027), 2 (0.081), 3 (0.119), 4 (0.054), 5 (0.086); palpal sensilla apparently restricted to middle third of palp 3. Cibarium with about 20 dot-like remnants of teeth, mostly at sides when viewed as in Fig. 10, no cibarial arch or pigment patch. Pharynx 0.19 long, unarmed. Pleura with 7-8 upper and 2 lower episternal setae. Length of wing vein sections: *alpha* (0.49), *beta* (0.24), *delta* (0.10), *gamma* (0.11). Length of femora, tibiae and basitarsi:

foreleg, 0.78, 1.30, 0.76; midleg, 0.71, 1.40, 0.93; hindleg, 0.83, 1.50, 1.02. Abdominal tergite 2 with ca. 20 erect hair sockets, tergite 3 with fewer, none visible on other tergites. Genitalia. Style 0.15 long, with 4 major spines, basal pair inserted slightly beyond middle of segment; no subterminal seta. Coxite 0.29 long, without persistent hairs. Paramere simple. Aedeagus subtriangular, well pigmented throughout, acute tip. Genital pump 0.116 long, each filament 0.35 or ca. 3X length of pump. Lateral lobe 0.24 long. Cereus as shown.

Female allotype. Wing length 1.98; width 0.56. Coloration as for ♂. Head height 0.47 (including 0.17 long clypeus); width 0.33. Eyes large, separated by 0.05 or by distance = to ca. 2.5 facet diameters. Interocular suture reduced to short stub. Flagellomere I 0.28 long, combined length of II + III = 0.24; ascoids as in ♂ but absent from terminal 2 flagellomeres. Labrum 0.28 long. Ventral maxillary teeth 25; lateral teeth 13. Length of palpal segments: 1 (0.035), 2 (0.116), 3 (0.159), 4 (0.060), 5 (0.110); palpal sensilla at middle third of segment. Cibarium with 9 horizontal pointed teeth, a few lateral teeth and ca. 36 ventral teeth, median ones larger than others; cibarial arch complete; pigment patch well infuscated, shaped as shown. Pharynx 0.21 long, unarmed. Pleura with 4 upper and 1 lower episternal setae. Length of wing vein sections: *alpha* (0.52), *beta* (0.26), *delta* (0.06), *gamma* (0.09). Length of femora, tibiae and basitarsi: foreleg, 0.83, 1.24, 0.80; midleg, 0.73, 1.47, 0.90; hindleg, 0.83, 1.50, 0.98. Abdominal tergite 2 with 30+ erect hair sockets, tergite 3 with 28, few or no erect setae on other tergites. Spermathecae

with 9-12 segments increasing in size from base to apex, terminal knob oblong, its length at least twice width; individual ducts nearly as long as common duct. Stem of genital fork very broad, blade-like.

Type Data (All material from Amazonas State, Brazil). Holotype ♂. 26 km E of Manaus at Reserva Ducke, 19 March 1979, flight trap, D.G. Young. Allotype ♀. 243 km E of Manaus at Rio Urubu, 13 March 1979, flight trap, D.G. Young, J.R. Arias et al. Paratypes. 2♀♀, 30 km E of Manaus, 3 Oct. 1974, rat-baited Disney trap, J.R. Arias. 3♂♂, Mauá, Estrada do Aleixo (km 10), Manaus, 18 Sept. 1974, lizard-baited Disney trap, J.R. Arias. 3♂♂, 18♀♀, 243 km E of Manaus at Rio Urubu, 12-14 March 1979, light and flight traps, human bait, D.G. Young, J.R. Arias et al. 8♂♂, 12♀♀, 26 km E of Manaus at Reserva Ducke, 16-19 March 1979, flight and light traps, D.G. Young, J. Vidal and R. Alves de Freitas. 2♂♂, 3♀♀, 43 km NE of Manaus at Reserva Campinas, 23 March 1979, light and flight traps, D.G. Young, J. Vidal and R. Alves de Freitas. The subspecific name, "nociva," meaning hurtful or injurious, refers to the bloodsucking habit of the females.

Discussion:

The *flaviscutellata* complex of the subgenus *Nyssomyia* Barretto now consists of 5 named and 1, or possibly 2, unnamed forms. These are: *Lu. flaviscutellata* (Mangabeira, 1942) from South America, *Lu. olmeca olmeca* (Vargas and Diaz-Najera, 1959) from Mexico and Central America, *Lu. o. bicolor* Fairchild and Theodor, 1971, from Costa Rica to northern South America, *Lu. o. nociva* n.ssp. from North Central Brazil, and *Lu. inornata* Martins, Falcão and da Silva, 1965, from Brazil. Specimens

from Salvador, Bahia, Brazil (Sherlock and Carneiro, 1962, not *flaviscutellata* Mang.) and from Itiatuba, Parâ, Brazil (Lewis, 1975) require further study before their status can be determined.

Lu. inornata, known from only the male, is structurally similar to *flaviscutellata* but the scutellum is dark rather than pale according to Martins et al. (1965). This feature, if consistent, also serves to separate this species from the subspecies of *olmeca*. Through the kindness of M. Emile Abonnenc and Prof. Nicole Leger, we examined a male, tentatively identified and recorded as *inornata* from Oyapock, French Guiana (Leger et al., 1977, Fig. 2). We believe, however, that the specimen is conspecific with *flaviscutellata* because the scutellum is paler than the mesonotum, palp 5 is shorter than palp 3, and the rounded tips of the genital filaments are similar to those of some *flaviscutellata* males from Pará, Brazil. These males, unlike those of *olmeca* and subspecies, have large genital pumps which are almost as long as the lateral lobes (Table 1).

Palpal segment 5 of *Lu. o. bicolor* and *Lu. o. nociva* males is subequal in length to palp 2; whereas palp 5 of *Lu. o. olmeca* is nearly as long as palp 3. The male of *nociva* is smaller than the other subspecies based on the length of the wings, flagellomeres, and lateral lobes of the genitalia. Also the narrow interocular distance of *nociva* males is useful in separating this subspecies from the others (Table 1).

Features of the spermathecae, cibarial armatures, palpi, and antennae are diagnostic for the *flaviscutellata* complex females (Fairchild and Theodor, 1971). The broad terminal knobs of the spermathecae of *Lu. o. olmeca* and the *flaviscutellata* complex female of Sherlock and Carneiro (1962) differ from the oblong knobs of the other females,

especially *nociva* (Figs. 10, 11, 12). Females of *nociva* and the nominate subspecies are distinguished from the others in the complex in having the individual and common sperm ducts subequal in length. The individual sperm ducts of the other females are markedly shorter than the common duct. Additional measurements and meristic characters for each taxon are shown in Table 1.

It should be noted that the broad, blade-like stem of the genital fork reaches its greatest development in *nociva* females. Not all are as wide as that shown in Fig. 7, but this is apparently due to viewing the structure at an improper angle.

The status of *Lu. o. nociva* may change after more is learned about its geographic distribution, especially in relation to that of *Lu. o. bicolor* which occurs in the Amazon basin at Leticia, Colombia, and Napo Province, Ecuador (Young, 1979). At present, there is no evidence indicating that the *olmeca* subspecies have sympatric distributions. *Lu. flaviscutellata*, however, does occur with two of the *olmeca* subspecies in Amazonia, supporting its position as a valid species and raising interesting questions about resource partitioning and disease relationships.

Acknowledgments

Mr. Alves de Freitas and Mr. João Ferreira Vidal collected and prepared many of the specimens. We also thank Prof. N. Leger and M. Emile Abonnenc for sending us phlebotomines from French Guiana for study.

Character \ Taxon	*Lu. flaviscutellata* ♂♂	*Lu. flaviscutellata* ♀♀	*Lu. inornata* ♂	*Lu. olmeca olmeca* ♂♂	*Lu. olmeca olmeca* ♀♀	*Lu. olmeca bicolor* ♂♂	*Lu. olmeca bicolor* ♀♀	*Lu. olmeca nociva* ♂♂	*Lu. olmeca nociva* ♀♀
nterocular istance	0.048-0.055 ($\bar{x}$ = 0.51) n = 10	0.050-0.068 ($\bar{x}$ = 0.62) n = 10	?	0.098-0.110 ($\bar{x}$ = 0.105) n = 10	0.106-0.113 ($\bar{x}$ = 0.109) n = 10	0.088-0.096 ($\bar{x}$ = 0.090) n = 10	0.099-0.101 ($\bar{x}$ = 0.952) n = 10	0.035-0.040 ($\bar{x}$ = 0.037) n = 8	0.050-0.060 ($\bar{x}$ = 0.053) n = 10
ength of lagello- ere II	0.29-0.32	0.30-0.33	0.28	0.53	0.37-0.42	0.48	0.31-0.38	0.31 n = 8	0.27-0.32 n = 13
umber of orizontal eeth in ibarium	--	6-7	--	--	10-12	--	8-9 (rarely 7 or 10)	--	9-12 (usually 10) n = 13
ength of enital ump	0.20	--	0.19	0.13-0.16	--	0.14	--	0.11 n = 8	--
ength of ateral obe	0.20	--	0.20	0.40	--	0.32	--	0.22-0.23 n = 8	--

ble 1. Measurements and counts of some features of the *flaviscutellata* complex species (from Fairchild and Theodor, 1971; Martins et al., 1965 for *Lu. inornata*; this paper where sample size (n) is given). Measurements in millimeters.

References

Aitken, T.H.G., J.P. Woodall, A.H.P. de Andrade, G. Bensabath and R.E. Shope. 1975. Pacui virus, phlebotomine flies and small mammals in Brazil: An epidemiological study. Amer. J. Trop. Med. Hyg. 24:358-368.

Disney, R.H.L. 1966. A trap for phlebotomine sandflies attracted to rats. Bull. Ent. Research 56:445-451.

Fairchild, G.B. and O. Theodor. 1971. On *Lutzomyia flaviscutellata* (Mangabeira) and *L. olmeca* (Vargas and Diaz-Najera) (Diptera: Psychodidae). J. Med. Ent. 8:153-159.

Lainson, R. and J.J. Shaw. 1968. Leishmaniasis in Brazil; II. Observations on enzootic rodent leishmaniasis in the lower Amazon region--The feeding habits of the vector, *Lutzomyia flaviscutellata* in reference to man, rodents, and other animals. Trans. R. Soc. Trop. Med. Hyg. 62:396-405.

_____. 1979. The role of animals in the epidemiology of South American leishmaniasis. In Biology of the Kinetoplastida, Vol. 2, Chapter 1 [W.H.R. Lumsden and D.A. Evans, eds.], Academic Press, London.

Leger, N., E. Abonnenc, F.X. Pajot, R. Kramer and J. Claustre. 1977. Liste commentée des phlébotomes de la Guyane Française. Cah. O.R.S.T.O.M. Ser. Ent. Med. Parasit. 15:217-232.

Lewis, D.J. 1975. The *Lutzomyia flaviscutellata* complex (Diptera: Psychodidae). J. Med. Ent. 12:363-368.

Mangabeira, O. 1942. Contribuição ao estudo dos flebotomus (Diptera: Psychodidae). Descrição dos machos de 24 novas espécies. Mem. Inst. Oswaldo Cruz 37:111-218.

Martins, A.V., A.L. Falcão and J.E. da Silva. 1965. Notas sôbre os flebótomos do Territorio de Rondônia, com a descrição de seis espécies novas (Diptera, Psychodidae). Rev. Brasil. Biol. 25:1-20.

Sherlock, I.A. and M. Carniero. 1962. Algumas fêmeas de *phlebotomus* do Brasil (Diptera, Psychodidae). Mem. Inst. Oswaldo Cruz 60:423-435.

Tikasingh, E.A. 1975. Observations on *Lutzomyia flaviscutellata* (Mangabeira) (Diptera: Psychodidae), a vector of enzootic leishmaniasis in Trinidad, West Indies. J. Med. Ent. 12:228-232.

Vargas, L. and A. Diaz-Nájera. 1959. *Phlebotomus farilli* n.sp., *Ph. humboldti* n.sp. y *Ph. olmecus* n.sp. de Mexico (Diptera, Psychodidae). Rev. Inst. Salubr. Enferm. Trop. 19:141-153.

Ward, R.D., R. Lainson and J.J. Shaw. 1973. Further evidence of the role of *Lutzomyia flaviscutellata* (Mangabeira) as the vector of *Leishmania mexicana amazonensis* in Brazil. Trans. R. Soc. Trop. Med. 67:608-609.

_____. 1977. Experimental transmissions of *Leishmania mexicana amazonensis* Lainson and Shaw, between hamsters by the bite of *Lutzomyia flaviscutellata* (Mangabeira). Trans. R. Soc. Trop. Med. Hyg. 71:265-266.

Young, D.G. 1979. A Review of the bloodsucking Psychodid Flies of Colombia (Diptera: Phlebotominae and Sycoracinae). Univ. of Florida Agr. Exp. Sta. Tech Bull. 806. 266 p.

Footnotes

1) Research was partly funded by CNPq Grant SIP 08/131; INPA Project 2017/103, and U.S. Army Medical Research and Development Contract DADA 17-72-C-2139.

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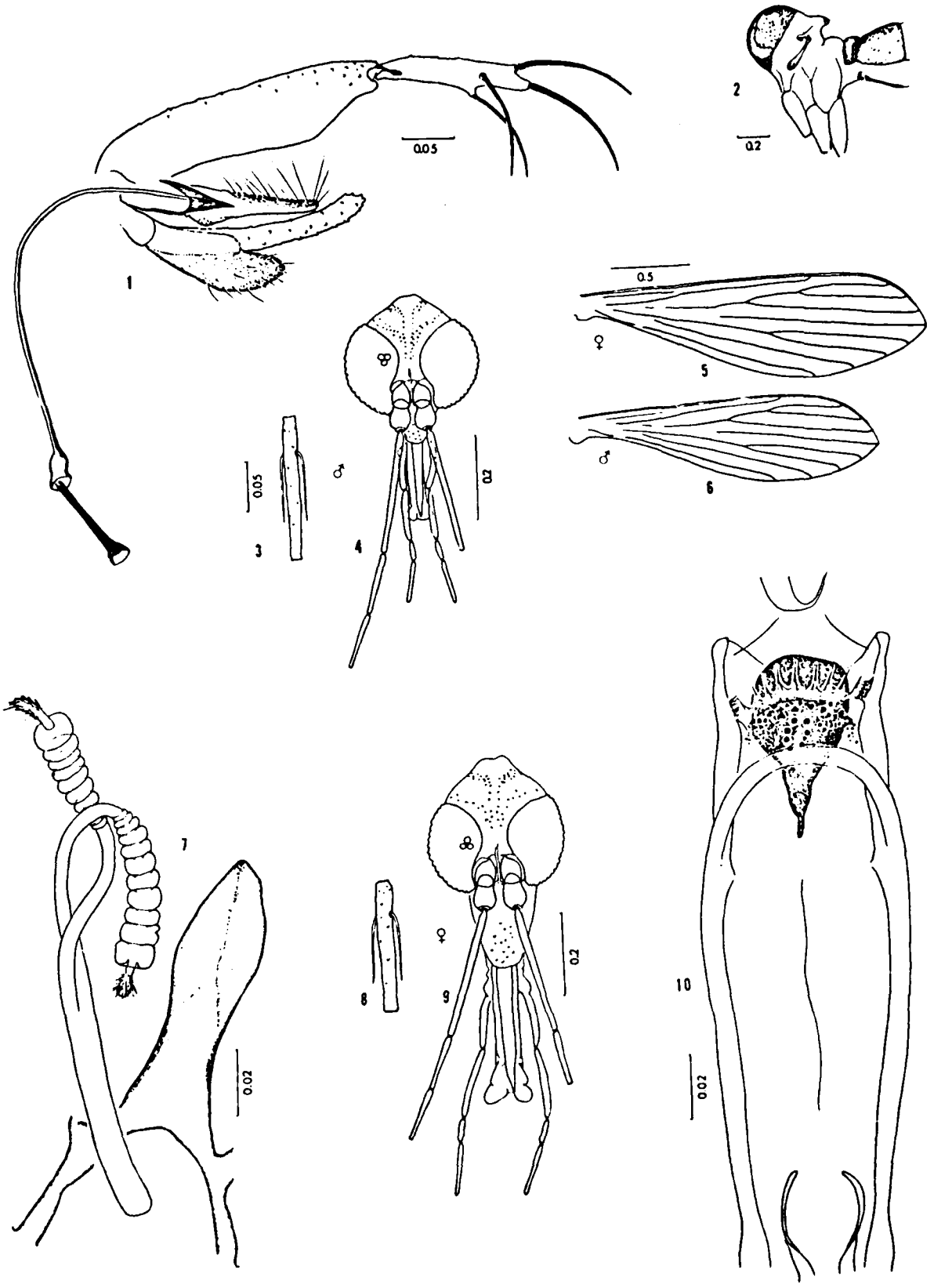
Figures

Figs. 1-10. *Lutzomyia olmeca nociva* n.ssp. 1) Holotype genitalia, lateral view. 2) Thorax and anterior abdomen of female. 3) Male flagellomere II. 4) Male head. 5) Female wing. 6) Male wing. 7) Spermathecae and genital fork. 8) Female flagellomere II. 9) Female head. 10) Female cibarium. Scale in mm.

Fig. 11. Spermathecae of *Lutzomyia flaviscutellata* of female collected with *Lu. olmeca nociva* at Rio Urubu, Brazil.

Fig. 12. Spermathecae of *Lutzomyia olmeca bicolor* (allotype). Figures 11 and 12 drawn at same scale as Fig. 7.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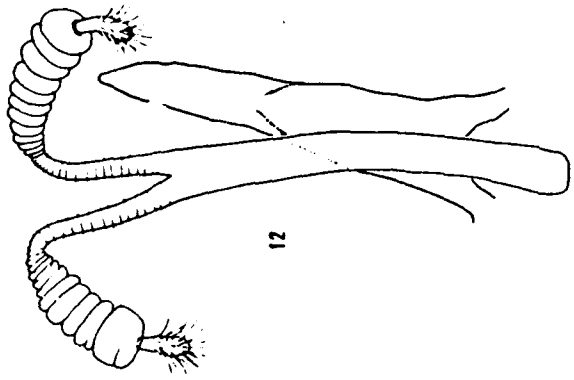

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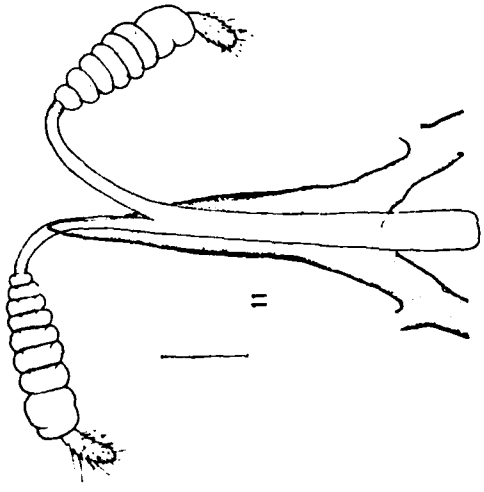

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Appendix IV

A Larval Diet for Rearing Phlebotomine Sand Flies (Diptera: Psychodidae)

Among the major problems in rearing phlebotomine sand flies is excessive larval mortality caused by fungal growth, improper diet or moisture, disease and/or other factors (Killick-Kendrick, R. 1978. Acta Tropica 35:297-313). None of the 600 or so species have been mass reared and only 6 or 7 have been reared in large numbers for more than 10 consecutive generations (Killick-Kendrick, op. cit.).

We have found that an aged 1:1 mixture (by wt. or vol.) of dry rabbit feces and Purina Rabbit Chow complete Diet #5315 (or Purina Horse Chow Checkers #3501) is an excellent diet for the larvae. The feces were obtained from laboratory rabbits fed on the Lab Rabbit Chow. Our attempts to rear four Lutzomyia spp. from the southern U.S.A. have been successful. We currently maintain closed colonies of Lutzomyia sp. near cruciata (Coq.), an autogenous strain (12 generations), L. anthophora (Addis) (6 generations) and L. shannoni (Dyar) (4 generations). We reared L. vexator (Coq.) in 1976 but purposely abandoned the colony after 4 generations.

The larval food is prepared by grinding the feces and whole chow into small pieces with a mortar and pestle. The final size of the particles ranges from 0.01 to 2.0 mm in diameter, the majority being about 0.5 to 1.0 mm. The dry mixture is evenly spread over the bottom of a petri dish or similar container to a depth of 0.5 to 1.0 cm, saturated

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[1]The chemical composition of these diets is available from the Ralston Purina Co., Checkerboard Square, St. Louis, Missouri 63188. Changes in composition during and after ageing due to microorganisms are not presently known.

with distilled water and placed in a glass desiccator at 22-30°C with 100% R.H. It is not sterilized. A crude inoculum of spores of the common bread mold, Rhizopus sp., is transferred from older diet to fresh using a wooden applicator. The desiccator is then covered with a tight fitting lid to prevent entry of phorid flies and other arthropods. The mixture is allowed to age for a month or more (Safyanova, V.M. 1964. Bull. Wld. Hlth. Org. 31:573-576). It is ready for use after most or all of the visible fungi disappears and when the odor becomes similar to that of rich humus. This ageing process eliminates the problem of excessive fungi which, by sheer density, can immobilize and kill larvae.

We rear 1 to 70 larvae in 7 dr (25 ml) polystrene snap-cap vials at temperatures ranging from 23-27°C and 80-100% R.H. The bottom half of each vial is filled with plaster of paris, allowed to dry, then saturated with distilled water. Gravid sand flies, usually 1 per vial, oviposit in these containers. The moist, aged medium is introduced into the vials anytime before egg hatch (about 1 g of medium for 50 eggs). As the larvae grow, we do not remove their feces but sometimes add more food or water. Little else is required for their care. Larval mortality, consistently less than 10%, results from factors other than improper nutrition such as accidents due to handling or excess water.

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LIST OF PUBLICATIONS RESULTING FROM THIS RESEARCH

1. Young, D.G. and C.H. Porter. 1972. *Luztomyia yuilli*, a new man-biting phlebotomine sand fly from Colombia (Diptera: Psychodidae). J. Med. Ent. 9(6):524-526.

2. Young, D.G. 1973. Two new phlebotomine sand flies from Colombia (Diptera: Psychodidae). Fla. Ent. 56(2):106-112.

3. Young, D.G. and C.H. Porter. 1974. *Lutzomyia cirrita* n. sp. from Colombia with a new synonym in the genus (Diptera: Psychodidae: Phlebotominae). Fla. Ent. 57(3):321-325.

4. Young, D.G. and D.J. Lewis. 1977. Pathogens of phlebotomine sand flies. Bull. W.H.O. (in press).

5. Young, D.G. and J.R. Arias. 1977. *Lutzomyia* sand flies in the subgenus *Evandromyia* Mangabeira with a description of a new species from Brazil. Acta Amazonica 7:59-70.

6. Lainson, R., R.D. Ward, D.G. Young, J.J. Shaw, and H. Fraiha. 1977. Preliminary entomological and parasitological studies in Humboldt, Aripuana, Mato Grosso State, Brazil. Acta Amazonica 6:55-60.

7. Lewis, D.J., D.G. Young, G.B. Fairchild, and D.M. Minter. 1977. Proposals for a stable classification of the phlebotomine sandflies (Diptera: Psychodidae). Syst. Ent. 2:319-332.

8. Aitken, T.H.G., A.J. Main, and D.G. Young. 1977. *Lutzomyia vexator* (Coquillett) in Connecticut (Diptera: Psychodidae). Proc. Ent. Soc. Wash. 79:582.

9. Young, D.G. 1979. A review of the bloodsucking psychodid flies of Colombia (Diptera: Phlebotominae and Sycoracinae). Bull. 806 (Technical) Agr. Exp. Sta. Univ. Florida. 266 p.

10. Young, D.G. and J.R. Arias. 1981. Sand flies of the Central Amazon of Brazil. 2. Description of *Lutzomyia (Trichophoromyia) ruii* n. sp. (Diptera: Psychodidae). Rev. Brasil. Biol. (in press).

11. Young, D.G., P.V. Perkins and R.G. Endris. 1981. A standard larval diet for rearing phlebotomine sand flies (Diptera: Psychodidae). J. Med. Ent. (submitted).

12. Abonnenc, E., J. Arias, N. Leger and D.G. Young. 1981. Sur *Lutzomyia davisi* (Root, 1934) et les especes de morphologie comparable (Diptera, Phlebotomidae). Ann. Parasit. Hum. Comp. (Review of the *Lutzomyia davisi* complex)(in press).

13. Young, D.G. and J.R. Arias. 1981. A new phlebotomine sand fly in the *Lutzomyia flaviscutellata* complex from northern Brazil (Diptera: Psychodidae). Rev. Brasil. Biol. (in press).

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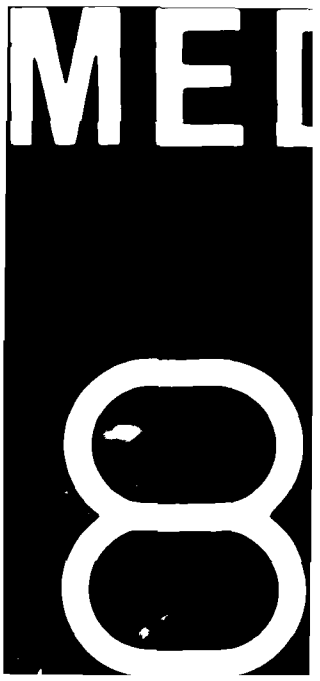
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